Absolute stereostructures of novel cytotoxic metabolites, gymnastatins A–E, from a *Gymnascella* species separated from a *Halichondria* sponge



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Gymnastatins A–E have been isolated from a strain of *Gymnascella dankaliensis* originally separated from the sponge *Halichondria japonica*, and their absolute stereostructures have been established on the basis of spectroscopic analyses using 1D and 2D NMR techniques and some chemical transformations. Among them gymnastatins A–C exhibited significant cytotoxicity against cultured P388 cells.

Introduction

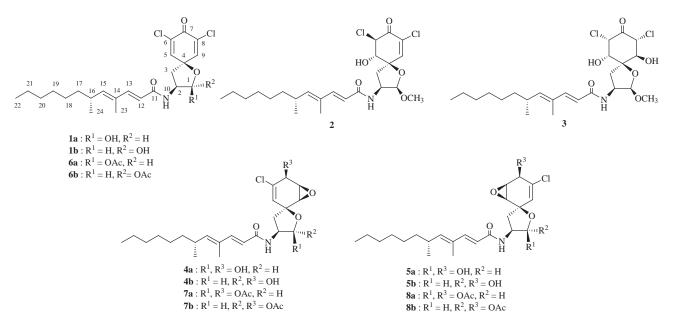
Based on the fact that some of the bioactive materials isolated from marine animals have been produced by bacteria,¹⁻⁴ we have focused our attention on new antitumour materials from microorganisms inhabiting the marine environment. As part of this program, we have found a number of antitumour and/or cytotoxic compounds from various fungi and an actinomycete originally isolated from various marine organisms, and elucidated their structures.⁵⁻¹² Our continuing search for cytotoxic compounds has led to the isolation of five metabolites designated gymnastatins A-E 1-5 from a strain of Gymnascella dankaliensis (Castellani) Currah OUPS-N134 which was separated from the sponge Halichondria japonica. We report herein the isolation and absolute stereostructure determination of 1-5. of which the relative stereostructures for 1-3 except for the configuration of the 16-position have been briefly reported in a preliminary form,¹³ together with their cytotoxic activities.

Results and discussion

The fungal strain was cultured at 27 $^{\circ}$ C for 4 weeks in a medium containing 1% malt extract, 1% glucose and 0.05% peptone in artificial seawater adjusted to pH 7.5. The MeOH extract of the

mycelia was purified by bioassay-directed fractionation employing a combination of Sephadex LH-20 and silica gel column chromatography and high-performance liquid chromatography (HPLC) to afford gymnastatins A–E **1–5** as colourless powders.

Gymnastatin A 1 had the molecular formula C₂₃H₃₁Cl₂NO₄ established by respective $[M - H_2O]^+$ and M^+ peaks of 1 and its acetate (6a) in high-resolution electron impact mass spectrometry (HREIMS), and the ratio of relative intensity of isotope peaks $(M^+:[M+2]^+:[M+4]^+ = ca. 9:6:1)$ in EIMS. Its IR spectrum exhibited bands at 3377, 3277, 1697, 1653 and 1606 cm⁻¹, characteristic of an alcohol, an amine, a conjugated ketone, an amide and a double bond. The ¹H and ¹³C NMR spectra (Table 1) of 1 suggested that it existed in a 2:1 mixture of two stereoisomers (1a and 1b), in which proton and carbon signals of two methines ($\delta_{\rm H}$ 5.54, d, J 4.3 Hz, and $\delta_{\rm H}$ 5.53, s; $\delta_{\rm C}$ 96.50 and $\delta_{\rm C}$ 103.58), assignable to a hemiacetal group, were observed. Two fractions separated from the mixture by HPLC each exhibited the NMR spectrum for a mixture of 1a and 1b in a 2:1 ratio. On the other hand, acetylation of 1 afforded two separable monoacetates (6a and 6b), in which the ¹H and ¹³C NMR signals for acetoxymethine groups appeared at $\delta_{\rm H}$ 6.38 (d, J 4.6 Hz) and $\delta_{\rm C}$ 95.24, and $\delta_{\rm H}$ 6.38 (s) and $\delta_{\rm C}$ 101.18, respectively (Table 2). Hydrolysis of 6a by an aqueous ammonia solution gave compound 1 as a 2:1 mixture of two stereoisomers.



	1a			1b		
Position	$\delta_{\mathbf{H}}{}^{a}$	J/Hz	$\delta_{ m C}$	$\overline{\delta_{\mathbf{H}}^{a}}$	<i>J</i> /Hz	$\delta_{\mathbf{C}}$
1	5.54d	4.3 (2)	96.50 (t) ^b	5.53s		103.58 (t)
2 3α	4.79m		52.24 (t)	4.65m		58.15 (t)
3α	2.59dd	12.9 (3 ^β), 8.3 (2)	38.35 (s)	2.82dd	$12.9(3\beta), 8.3(2)$	40.21 (s)
β	2.24t	12.9 (2,3α)		2.24t	12.9 (2,3α)	
4			78.98 (q)			80.30 (q)
5	7.03d	2.7 (9)	144.72 (t)	7.03d	2.7 (9)	144.72 (t)
			130.61 (q)			130.61 (q)
7			172.78 (q)			172.78 (q)
6 7 8			130.69 (q)			130.69 (q)
9	7.14d	2.7 (5)	147.27 (t)	7.14d	2.7 (5)	147.27 (t)
10	6.18d	8.2 (2)		6.18d	8.2 (2)	
11			166.99 (q)			167.78 (q)
12	5.78d	15.1 (13)	116.64 (t)	5.78d	15.1 (13)	116.64 (t)
13	7.26d	15.1 (12)	147.84 (t)	7.26d	15.1 (12)	147.84 (t)
14			130.80 (q)			130.80 (q)
15	5.78d	9.8 (16)	148.98 (t)	5.78d	9.8 (16)	148.98 (t)
16	2.50m		33.24 (t)	2.50m		33.24 (t)
17	1.37m		37.21 (s)	1.37m		37.21 (s)
18	1.23m		27.48 (s)	1.23m		27.48 (s)
19	1.23m		29.39 (s)	1.23m		29.39 (s)
20	1.23m		31.83 (s)	1.23m		31.83 (s)
21	1.23m		22.64 (s)	1.23m		22.64 (s)
22	0.87t	6.7 (21)	14.11 (p)	0.87t	6.7 (21)	14.11 (p)
23	1.77s		12.52 (p)	1.77s		12.52 (p)
24	0.96d	6.6 (16)	20.51 (p)	0.96d	6.6 (16)	20.51 (p)
1-OH	5.11br s		···· (1)	5.11br s		··· ··· ··· ··· ··· ··· ··· ··· ··· ··

^{*a*}¹H Chemical-shift values (δ /ppm from TMS) followed by multiplicity and then the coupling constant (*J*/Hz). Figures in parentheses indicate the proton coupling with that position. ^{*b*} Letters, p, s, t and q, in parentheses indicate, respectively, primary, secondary, tertiary and quaternary carbons, assigned by DEPT.

This evidence implied that 1 contains two stereoisomers (1a and 1b) on a hemiacetal group, which coexist at equilibrium in a certain ratio. Acetate 6a thus obtained was used for a structure analysis of 1.

A close inspection of the ¹H and ¹³C NMR spectral data (Table 2) of **6a** by DEPT and ¹H-¹H and ¹H-¹³C correlation spectroscopy (COSY) experiments revealed the presence of the following functionalities: three methyls (C-22 to C-24) including one primary, secondary and vinyl methyl each, six methylenes (C-3 and C-17 to C-21), one disubstituted and three trisubstituted double bonds (C-12, C-13, C-5, C-6, C-8, C-9, C-14 and C-15), a secondary amide (N-10 and C-11) and a quaternary sp³-carbon (C-4) linked to an oxygen atom, two sp³-hybridized methines (C-2 and C-16) including one methine linked to a nitrogen atom in addition to the acetoxymethine (C-1) of the hemiacetal group. The remaining functionality, corresponding to the carbon signal at δ_c 172.08, was shown to be a ketone in a cross-conjugated dienone system on the basis of HMBC correlations (5-H/C-7 and 9-H/C-7) shown in Fig. 1. The carbon signal of the conjugated ketone appeared shifted upfield by ca. 10 ppm, relative to a general cross-conjugated cyclohexadienone ($\delta_{\rm C}$ 183–185),^{14–16} suggesting that chlorine atoms exist at the α -position of the ketone.^{16,17} In addition to the partial structure (C-5 to C-9) thus established, the ¹H-¹H COSY analysis of **6a** led to other partial structures shown by bold-faced lines in Fig. 1, which were supported by HMBC correlations (Table 2). The geometry of the diene in the side chain was deduced from a coupling constant $(J_{12,13} 15.1 \text{ Hz})$ of olefinic protons, a chemical shift ($\delta_{\rm C}$ 12.49) of the ¹³C NMR signal of a vinyl methyl,¹⁸ and nuclear Overhauser enhancements (NOEs) for 12-H/23-H and 13-H/15-H. The connection of these partial structures and the remaining functional groups (C-4, C-11 and C-18-C-20) was determined on the basis of HMBC correlations shown in Fig. 1. Based on this evidence, the planar structure of acetate 6a was elucidated. The side chain was supported by an EIMS fragment at m/z 179 $([C_{13}H_{23}]^+).$

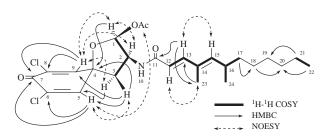


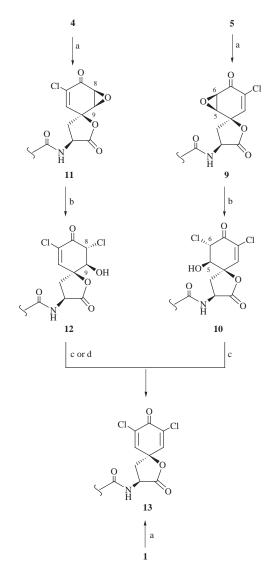
Fig. 1 Selected ${}^{1}H-{}^{1}H$ COSY, HMBC and NOESY correlations in gymnastatin A acetate 6a.

In an NOESY experiment, acetate **6a** showed NOEs for 1-H/ 9-H, 2-H/9-H and Ac/5-H (Fig. 1), whereas stereoisomeric acetate **6b** exhibited NOEs for 1-H/5-H, 2-H/9-H and Ac/9-H (Table 2). This finding indicated that 1-H and 2-H are in the same arrangement as 9-H in **6a**, and 1-OAc and 2-H are in the same arrangement as 9-H in **6b**, meaning that H-1 is *trans* to 2-H. Consequently, the relative stereostructures for gymnastatin A **1** as a mixture of diastereomers at the 1-position were established as **1a** and **1b** except for the configuration of the 16-position. The absolute stereostructure of **1** including the configuration of the 16-position was determined by a chemical transformation of gymnastatin E **5**, of which the absolute configuration was determined as described below, to the oxidation product **13** of **1** by a pyridine–CrO₃ complex (Scheme 1).

Gymnastatin E **5** was assigned the molecular formula $C_{23}H_{34}$ -ClNO₅ as deduced from a molecular ion peak in HREIMS. Its IR spectrum exhibited absorption bands for hydroxy and amino groups, an amide and a double bond, but did not show a band for a conjugated ketone. The ¹H and ¹³C NMR spectra (Table 3) of **5** suggested that it exists in a 2:1 mixture of two stereoisomers (**5a** and **5b**) as in compound **1**. Acetylation of **5** by a standard method gave two diacetates **8a** and **8b**. Since these acetates could be separated by HPLC, acetate **8a** was used for a structure analysis of **5**.

Position	-						6b			
obition	$\delta_{\rm H}{}^a$	J/Hz	$\delta_{\rm C}$	¹ H– ¹ H COSY	HMBC (C)	NOESY	$\delta_{\mathrm{H}}{}^{a}$	<i>J</i> /Hz	$\delta_{\rm C}$	NOESY
1	6.38d	4.6 (2)	95.24 (t) ^b	2	1-OCOCH ₃ , 2, 3, 4	2, 9, 10	6.38s		101.18 (t)	2, 3β, 5, 10
2	5.06dddd	$13.0(3\beta), 8.7(10), 7.8(3\alpha), 4.6(1)$	50.99 (t)	$1, 3\alpha, 3\beta, 10$		1, 3α, 9, 10	4.74td	$6.2(3\alpha, 10), 2.7(3\beta)$	56.78 (t)	1, 3α, 3β, 9, 1
3α	2.63dd	13.0 (3β), 7.8 (2)	38.42 (s)	3β	1, 2, 5	2, 3β, 9	2.75dd	14.4 (3β), 6.2 (2)	40.78 (s)	3β, 2, 9
β	2.20t	$13.0(2, 3\alpha)$		2, 3α	2, 4, 5, 9	3α, 5, 10	2.37dd	$14.4(3\alpha), 2.7(2)$		2, 3α, 5, 10
4			79.87 (q)	,	7 7 7 7				81.74 (q)	, , - , - ,
5	6.89d	2.7 (9)	145.03 (t)	9	6, 7, 9	3β, 1-OAc	6.99d	2.7 (9)	144.00 (t)	1, 3β
6			131.70 (q)	-	-, . , .				131.16 (q)	-,
7			172.08 (q)						172.11 (q)	
8			131.35 (q)						131.42 (q)	
-	7.05d	2.7 (5)	143.15 (t)	5	5, 7, 8	$1, 2, 3\alpha$	7.01d	2.7 (5)	145.28 (t)	1-OAc, 2, 3α
,	5.64d	8.7 (2)	115.15 (t)	2	2, 11	1, 2, 30 $1, 2, 3\beta$	5.85d	6.2 (2)	115.20 (1)	$1, 2, 3\beta$
1	5.0 4 0	0.7 (2)	166.26 (g)	2	2, 11	1, 2, 5p	5.650	0.2 (2)	166.57 (q)	1, 2, 5p
	5.73d	15.1 (13)	116.07 (t)	13	11, 13, 14	23	5.78d	15.2 (13)	116.08 (t)	23
	7.30d	15.1 (12)	148.37(t)	12	11, 12, 14, 15, 23	15	7.30d	15.2 (12)	148.34(t)	15
4	7.50 u	13.1 (12)	130.65 (q)	12	11, 12, 11, 10, 25	15	7.50 u	13.2 (12)	130.69 (q)	15
5	5.71d	9.6 (16)	149.20 (t)	16, 23	13, 14, 16, 17, 23, 24	13, 16, 24	5.70d	10.0 (16)	149.28 (t)	13, 24
	2.52m)10 (10)	33.28 (t)	15, 17A, 17B, 24	10, 11, 10, 17, 20, 21	15, 23, 24	2.52m	1010 (10)	33.29 (t)	23, 24
7A	1.27m		37.20 (s)	16	18	13, 23, 21	1.27m		37.21 (s)	23, 21
	1.35		57.20 (3)	16	18		1.35m		57.21 (5)	
8	1.22m		27.46 (s)	10	19		1.22m		27.47 (s)	
9	1.23m		29.37 (s)		19		1.24m		29.38 (s)	
20	1.27m		31.81 (s)		18		1.27m		31.81 (s)	
21	1.23m		22.60 (s)	22	19, 20		1.23m		22.62 (s)	
	0.87t	6.6 (21)	14.06 (p)	21	20, 21		0.87t	6.9 (22)	14.09 (p)	
	1.78s	0.0 (21)	12.49 (p)	15	13, 14, 15	12, 16	1.79s	0.9 (22)	12.51 (p)	12, 16
	0.97d	6.7 (16)	20.48 (p)	16	15, 16, 17	15, 16	0.98d	6.6 (16)	20.49 (p)	15, 16
-0 <i>C</i> OCH ₃	0.274	0.7 (10)	169.03 (q)	10	10, 10, 17	10,10	0.700	0.0 (10)	169.12 (g)	10, 10
	2.21s		21.34 (p)		1-0 <i>C</i> 0CH ₃	5	2.17s		21.18 (p)	9
,b As in Table 1			21.5 ((p)			5	2.175		21.10 (P)	/

Table 2 ¹H and ¹³C NMR data of gymnastatin A acetates 6a and 6b in CDCl₃



Scheme 1 Reaction conditions: (a) pyridine– CrO_3 ; (b) pyridinium chloride; (c) Ac_2O , pyridine; (d) MsCl, Et_3N .

The general features of the ¹H and ¹³C NMR spectra (Table 4) of **8a** closely resembled those of **6a** except that the signals for the ketone and one of the trisubstituted double bonds in the cyclohexane ring of **6a** were respectively replaced by those of a secondary alcohol ($\delta_{\rm H}$ 5.68; $\delta_{\rm C}$ 66.66) and an epoxide [$\delta_{\rm H}$ 3.34 and 3.58; $\delta_{\rm C}$ 56.88 ($J_{\rm CH}$ 183.6 Hz) and 51.48 ($J_{\rm CH}$ 180.2 Hz)] in **8a**. The position of the three functional groups in the cyclohexane ring of **8a** was established by a combination of coupling relationships between vicinal protons and ¹H–¹H COSY correlations from 6-H to 5-H and 7-H, and from 9-H to 7-H and 5-H (long-range couplings). It is most likely that a cyclohexane ring with an epoxide and a double bond like **8a** exists in a boat

conformation. The observation of NOEs from 9-H (a vinyl proton) to 1-H, 2-H and 3-H^{α}, and from 5-H (an epoxymethine proton) to 3-H^{β} in **8a** (Table 4) implied that the stereostructure of 8a is either of two stereoisomers, in which the orientation of the epoxide is opposite each other. In order to choose between them for 8a, gymnastatin E 5 was converted to keto-lactone 9, which then was derivatized to chlorohydrin 10 by treatment with pyridinium chloride (Scheme 1). The large coupling constant ($J_{5.6}$ 10.4 Hz) between H-5 and H-6 in chlorohydrin 10 obtained indicated that the cyclohexane ring of 10 exists in a chair conformation, with 5-OH and 6-Cl in a coequatorial arrangement. In NOESY experiments of compound 10, NOEs from 3-H^{α} to 2-H and 9-H (a vinyl proton), and from 3-H^{β} to 5-H (a hydroxymethine proton) were observed, implying that 5-OH in chlorohydrin 10 is oriented *cis* to the carbonyloxy group, and consequently the epoxide bond in acetate 8a is oriented cis to the ether bond (C-4-O). The summary of these considerations led to the relative stereostructures of acetate 8a and, consequently, of 5 (5a and 5b) for gymnastatin E, except for the 7- and 16-positions. In order to determine the relative configuration of the 7- and 16-positions in 5, an X-ray crystal structure analysis was carried out for a single crystal of acetate 8a (obtained by recrystallization from MeOH). The result obtained (Fig. 2) allowed assignment of the relative configuration of all the asymmetric centres and the conformation for 8a and 5. Though the chirality of 8a was also investigated by using the abnormal dispersion factor of chlorine in the X-ray analysis, a conclusive result was not obtained. Therefore, the modified Mosher method¹⁹ was applied for determination of the absolute configuration of compound 5.

The '(R)- and (S)-2-methoxy-2-phenyl-2-(trifluoromethyl)acetic acid' (MTPA) esters 14a and 14b of compound 5 were prepared by a standard method. Though compound 5 is a mixture of two stereoisomers (5a and 5b), this reaction gave only the 1,7-bis-MTPA esters with the 1-ester in a β -configuration, implying that these reaction conditions displaced the equilibrium of the hemiacetal group to one stereoisomer. The ¹H chemical-shift differences between the (R)- and (S)-1,7-bis-MTPA esters 14a and 14b are shown in Fig. 3, and the result suggested the R configuration for the asymmetric centre at C-7 and, consequently, led to absolute stereostructure 5 (5a and 5b) for gymnastatin E. It is known that ¹H chemical-shift differences of MTPA esters with axial arrangements or with steric hindrances are irregularly distributed.¹⁹ In this case, the 1-MTPA ester with steric hindrance also did not give data in accord with the rule.

As described above briefly, treatment of chlorohydrin 10 from 5 with acetic anhydride in pyridine afforded keto-lactone 13 which was identical with that derived from 1 by treatment with a pyridine– CrO_3 complex in all the respects including a specific rotation. This evidence led to the absolute configuration of all the asymmetric centres including the 16-position for gymnastatin A 1 (Scheme 1).

Gymnastatin D 4 is isomeric with compound 5. The general features of 1 H and 13 C NMR spectra (Table 5) of 4 closely

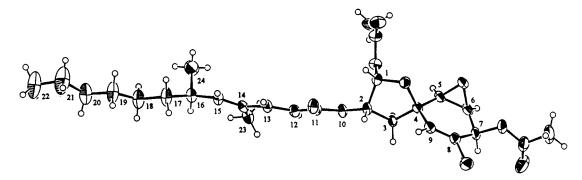
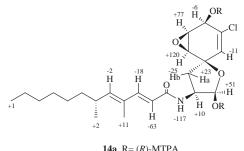


Fig. 2 X-Ray crystal structure for compound 8a.

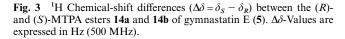
Table 3	¹ H and ¹	³ C NMR	data of	gymnastatin	E 5 in	acetone-d ₆
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	5a			5b		
Position	$\overline{\delta_{\mathrm{H}}{}^{a}}$	J/Hz	$\delta_{\rm C}$	$\delta_{\rm H}{}^a$	J/Hz	$\delta_{\mathbf{c}}$
1	5.42t	3.8 (2, 1-OH)	$96.65(t)^{b}$	5.48d	3.4 (1-OH)	103.13 (t)
2	4.60m		52.81 (t)	4.40m		58.09 (t)
3α	2.24dd	$12.4(3\beta), 8.2(2)$	39.38 (s)	2.44dd	$14.0(3\beta), 6.7(2)$	40.47 (s)
β	2.13t	$12.4(2,3\alpha)$		2.18t	$14.0(3\alpha), 1.4(2)$	
4			81.23 (q)			82.99 (q)
4 5	3.43dd	4.1 (6), 2.4 (9)	59.00 (t)	3.45dd	4.1 (6), 2.4 (9)	57.74 (t)
6	3.53dd	4.1 (5), 2.8 (7)	55.03 (t)	3.52dd	4.1 (5), 2.8 (7)	54.74 (t)
7	4.41dd	2.8 (6), 1.6 (9)	66.65 (t)	4.43dd	2.8 (6), 1.6 (9)	66.52 (t)
			132.58 (q)			132.15 (q)
8 9	5.80dd	2.4 (5), 1.6 (7)	129.08 (t)	5.86dd	2.4 (5), 1.6 (7)	131.19 (t)
10	7.13d	8.0 (2)		7.49d	6.6 (2)	
11			166.36 (q)			166.41 (q)
12	6.16d	15.4 (13)	119.99 (t)	6.11d	15.4 (13)	119.78 (t)
13	7.21d	15.4 (12)	145.74 (t)	7.23d	15.4 (12)	145.83 (t)
14			132.25 (q)			132.15 (q)
15	5.66d	9.8 (16)	146.66 (t)	5.66d	9.8 (16)	146.86 (t)
16	2.58m		33.68 (t)	2.58m		33.68 (t)
17A	1.29m		38.03 (s)	1.29m		38.03 (s)
В	1.38m			1.38m		()
18	1.26m		28.18 (s)	1.26m		28.18 (s)
19	1.27m		30.05 (s)	1.27m		30.05 (s)
20	1.26m		32.54 (s)	1.26m		32.54 (s)
21	1.27m		23.24 (s)	1.27m		23.24 (s)
22	0.89t	6.9 (21)	14.29 (p)	0.89t	6.9 (21)	14.29 (p)
23	1.81s	()	12.70 (p)	1.79s	()	12.70 (p)
24	1.00d	6.6 (16)	20.91 (p)			20.91 (p)
1-OH	6.05d	4.1 (1)	(1)	5.88d	3.7(1)	······································
7-OH	4.76d	8.5 (7)		4.79d	8.5 (7)	



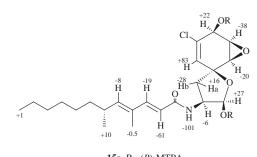


14a R= (*R*)-MTPA **14b** R= (*S*)-MTPA



resembled those of its isomer **5** and also indicated it to be a mixture of two stereoisomers (**4a** and **4b**) on the hemiacetal group. Moreover, compound **4** gave two separable diacetates **7a** and **7b**, of which the NMR spectra (Table 6) also exhibited signals similar to diacetates **8a** and **8b** of compound **5**, respectively.

In NOESY experiments of diacetate 7a, NOEs from 9-H (an epoxymethine proton) to 2-H and 3-H^{α}, and from 5-H (a vinyl proton) to $3-H^{\beta}$ were observed, whereas an NOE was not observed between 1-H and 9-H (Table 6). Though an NOE between 1-H and 2-H was observed in both diacetates 7a and 7b, there were obvious differences between the two acetates in selected difference NOE values between 1-H and 2-H [8.0% (7a) > 3.3% (7b)]. This finding suggested that 1-H and 2-H are oriented on the same side as the epoxymethine proton (9-H) in compound 7a. However, the orientation of the epoxide in 7a was not suggested by these observed NOEs. It was deduced from NOE data of chlorohydrin 12 derived by oxidation of compound 4 followed by treatment with pyridinium chloride as in the case of 5 (Scheme 1). The observation of NOEs from 5-H (a vinylic proton) to $3-H^{\beta}$, and from $3-H^{\alpha}$ to 2-H and 9-H (a hydroxymethine proton) in 12 implied that the epoxide bond in



15a R= (*R*)-MTPA **15b** R= (*S*)-MTPA

Fig. 4 ¹H Chemical-shift differences $(\Delta \delta = \delta_s - \delta_R)$ between the (*R*)and (*S*)-MTPA esters **15a** and **15b** of gymnastatin D (**4**). $\Delta \delta$ -Values are expressed in Hz (500 MHz).

acetate **7a** is oriented *cis* to the ether bond (C-4–O). The coupling constant (2.8 Hz) between 7-H and 8-H in **7a** was almost the same value as that between 6-H and 7-H in **8a**, suggesting the pseudoequatorial arrangement of 7-OH in **7a** as in **8a**. The above-summarized evidence led to the relative stereostructure of **4** (**4a** and **4b**) for gymnastatin D.

Treatment of chlorohydrin 12 with acetic anhydride in pyridine or mesyl chloride (MsCl) and triethylamine (Et₃N) afforded keto-lactone 13 derived from gymnastatin E 5 (Scheme 1), allowing assignment of the absolute configuration to gymnastatin D 4. This absolute configuration was identical with that assigned by application of the modified Mosher method for the (R)- and (S)-1,7-bis-MTPA esters 15a and 15b derived from 4 (Fig. 4). This result supported the pseudoequatorial arrangement of 7-OH in 4, deduced from the observed coupling constant between 7-H and 8-H in acetate 7a.

Gymnastatin B 2 had the molecular formula $C_{24}H_{35}Cl_2NO_5$ established by an M⁺ peak of 2 in HREIMS. The general spectral features of compound 2 closely resembled those of 1a except that one of the trisubstituted double bonds in the cyclohexane ring and the hydroxy group on the hemiacetal group in 1a were replaced by two sp³-methines linked to a hydroxy group

	8a						8b		
Position	$\delta_{\mathrm{H}}{}^{a}$	<i>J</i> /Hz	$\delta_{\rm C}$	¹ H– ¹ H COSY	HMBC (C)	NOESY	$\delta_{\mathrm{H}}{}^{a}$	J/Hz	$\delta_{\rm C}$
1	6.37d	4.4 (2)	$95.34(t)^{b}$	2	2, 3, 4	2, 9, 10	6.33s		100.55 (t)
2	4.94dddd	$12.5(3\beta), 9.4(10), 8.3(3\alpha), 4.4(1)$	50.85 (t)	$1, 3\alpha, 3\beta, 10$, ,	$1, 3\alpha, 9, 10$	4.63td	$6.5(3\alpha, 10), 1, 6(3\beta)$	56.46 (t)
3α	2.42dd	12.5 (3β), 8.3 (2)	38.89 (s)	2, 3β	2, 4, 5, 9	2, 3β, 9	2.48dd	14.3 (3β), 6.5 (2)	40.07 (s)
β	2.03t	$12.5(2, 3\alpha)$	()	2, 3α	1, 2, 5, 9	3a, 5	2.31dd	14.3 (3a), 1.6 (2)	
4			81.88 (q)	,	, , , , .	, -			83.72 (q)
5	3.34dd	4.1 (6), 2.3 (9)	56.88 (t)	9	4, 9	3β, 6	3.42dd	4.1 (6), 2.5 (9)	56.19 (t)
6	3.58dd	4.1 (5), 2.7 (7)	51.48 (q)	5, 7	8	5,7	3.57dd	4.1 (5), 2.7 (7)	51.58 (t)
7	5.68dd	2.7 (6), 1.8 (9)	66.66 (t)	6, 9	6, 9	6	5.68dd	2.7 (6), 1.8 (9)	66.52 (t)
8		(1),(2)	127.97 (q)	-,-	-,-			(1),(1)	127.73 (g)
9	5.88dd	2.3 (5), 1.8 (7)	128.68 (t)	5, 7	5, 7, 8	1, 2, 3α	5.81dd	2.5 (5), 1.8 (7)	129.94 (t)
10	5.69d	9.4 (2)	120100 (0)	2	5, 7, 6	1, 2, 50	5.94br d	6.5 (2)	1200 (0)
11	01074	··· (-)	166.23 (q)	-		-, -	bis for a	010 (2)	166.35 (q)
12	5.71d	15.2 (13)	116.36 (t)	13	11, 13, 14	23	5.73d	15.1 (13)	116.48 (t)
13	7.28d	15.2 (12)	147.98 (t)	12	11, 12, 14, 15, 23	15	7.26d	15.1 (12)	147.75 (t)
14	/120 G	10.2 (12)	130.65 (q)		11, 12, 11, 10, 20	10	/120 u	1011 (12)	130.73 (q)
15	5.59d	9.4 (16)	148.85 (t)	16, 23	13, 14, 16, 17, 23	13, 24	5.67d	9.6 (16)	148.81(t)
16	2.51m)(IO)	33.26 (t)	15, 17A, 17B, 24	10, 11, 10, 17, 20	23, 24	2.51m)10 (10)	33.24 (t)
17A	1.26m		37.22 (s)	16	18	20,21	1.25m		37.22 (s)
В	1.35m		0 / 122 (0)	16	18		1.35m		5 / 122 (0)
18	1.22m		27.47 (s)	10	19		1.20m		27.47 (s)
19	1.24m		29.38 (s)		.,		1.23m		29.38 (s)
20	1.23m		31.82 (s)		18		1.27m		31.82 (s)
21	1.27m		22.62 (s)	22	19, 20		1.23m		22.62 (s)
22	0.87t	6.9 (21)	14.08 (p)	21	20, 21		0.87t	6.9 (21)	14.09 (p)
23	1.77s	0.0 (21)	12.50 (p)	15	13, 14, 15	12, 16	1.76s	000 (21)	12.48 (p)
24	0.98d	6.6 (16)	20.51 (p)	16	15, 16, 17	15, 16	0.97d	6.6 (16)	20.51 (p)
1-OCOCH ₃		()	169.11 (q)		,,,		5.57.2		169.11 (g)
1-OCO <i>CH</i> ₃	2.17s		21.45 (p)		1-OCOCH3		2.11s		21.17 (p)
7-OCOCH ₃	2.1.75		170.42 (q)				2.1.10		170.34 (g)
7-OCO <i>CH</i> ₃	2.21s		20.71 (p)		7-OCOCH ₃		2.21s		20.69 (p)
^{<i>a,b</i>} As in Table			2000 f (p)		,		2.210		(P)

 Table 4
 ¹H and ¹³C NMR data of gymnastatin E diacetates 8a and 8b in CDCl₃

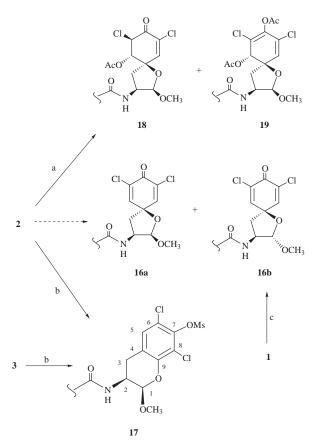
Table 5	¹ H and ¹³ C N	AR data of	f gymnastatin	D (4) in acetone- d_6
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		4a			4b		
	Position	$\delta_{\mathbf{H}}{}^{a}$	<i>J</i> /Hz	$\delta_{ m c}$	$\overline{\delta_{\mathbf{H}}^{a}}$	J/Hz	$\delta_{\rm C}$
	1	5.40t	4.1 (2, 1-OH)	96.64 (t) ^b	5.46d	3.7 (1-OH)	103.13 (t)
	2	4.62m		52.64 (t)	4.42m		57.73 (t)
	3α	2.44dd	$12.6(3\beta), 8.0(2)$	38.33 (s)	2.63dd	$14.0(3\beta), 6.8(2)$	40.80 (s)
	β	1.99dd	$12.6(3\alpha), 11.4(2)$		1.93dd	$14.0(3\alpha), 2.1(2)$	
	4			80.86 (q)			82.83 (q)
	5	5.76dd	2.3 (9), 1.6 (7)	130.47 (t)	5.79dd	2.3 (9), 1.6 (7)	129.54 (t)
	6			132.18 (g)			132.08 (q)
	7	4.40dd	2.9 (8), 1.6 (5)	66.47 (t)	4.42dd	2.5 (9), 1.6 (5)	66.53 (t)
	8	3.55dd	4.1 (9), 2.9 (7)	54.79 (t)	3.53dd	4.1 (9), 2.9 (7)	55.13 (t)
	9	3.43dd	4.1 (8), 2.3 (5)	57.46 (t)	3.48dd	4.1 (8), 2.3 (5)	59.15 (t)
	10	7.17d	8.0 (2)		7.52d	6.6 (2)	
	11			166.67 (q)			166.67 (q)
	12	6.15d	15.3 (13)	119.77 (t)	6.05d	15.3 (13)	119.56 (t)
	13	7.21d	15.3 (12)	145.98 (t)	7.21d	15.3 (12)	145.98 (t)
	14			132.22 (q)			132.22 (q)
	15	5.64d	9.8 (16)	146.95 (t)	5.64d	9.8 (16)	146.95 (t)
	16	2.57m		33.65 (t)	2.57m		33.65 (t)
	17A	1.29m		37.96 (s)	1.29m		37.96 (s)
	В	1.38m			1.38m		()
	18	1.29m		28.14 (s)	1.29m		28.14 (s)
	19	1.28m		30.05 (s)	1.28m		30.05 (s)
	20	1.29m		32.49 (s)	1.29m		32.49 (s)
	21	1.26m		23.20 (s)	1.26m		23.20 (s)
	22	0.87t	6.9 (21)	14.28 (p)	0.87t	6.9 (21)	14.28 (p)
	23	1.79s		12.69 (p)	1.76s		12.69 (p)
	24	0.99d	6.7 (16)	20.88 (p)	0.99d	6.7 (16)	20.88 (p)
	1-OH	6.05d	4.1 (1)	(L)	5.88d	3.7 (1)	
	7-OH	4.76d	8.5 (7)		4.79d	8.5 (7)	
in Table 1.							

and a chlorine atom $[\delta_{\rm H} 4.29 \ (5-{\rm H}), \delta_{\rm C} 75.46 \ ({\rm C}-5); \delta_{\rm H} 5.40 \ (6-{\rm H}), \delta_{\rm C} 60.85 \ ({\rm C}-6)]$, and a methoxy group $(\delta_{\rm H} 3.48; \delta_{\rm C} 55.10)$ in **2**, respectively, and the C-7 signal $(\delta_{\rm C} 183.49)$ of the ketone in **2** appeared shifted downfield relative to that of **1a** in the NMR spectra (Table 7). The planar structure of **2** thus deduced from the NMR spectral analysis was confirmed by analysis of ¹H⁻¹H COSY and HMBC correlations (1-H/OMe, 5-H/C-4, 5-H/C-7, 5-H/C-9, 9-H/C-3, 9-H/C-7 *etc.*; Table 7).

The observations of NOEs from 9-H to 2-H and $3-H^{\alpha}$, and from 5-H to $3-H^{\beta}$ and OMe, and a W-type long-range coupling $(J_{5,9}, 2.2 \text{ Hz})$ between 5-H and 9-H in 2 (Table 7) showed that 1-H and 2-H are on the same side as 9-H, and the 1-methoxy group is on the same side as 5-H with a pseudoequatorial arrangement on the cyclohexane ring. In addition, observation of NOEs from 6-H to 5-H and 5-OH, and the small coupling constant $(J_{5,6}, 2.2 \text{ Hz})$ between 5-H and 6-H implied that the cyclohexane ring in 2 exists in a twist-chair conformation with 5-H and 6-H in a co-pseudoequatorial, consequently, *trans* arrangement.

An interconversion of compounds 2 and 1 was attempted for assignment of an absolute configuration for 2. Treatment of 1 with trimethyl orthoformate under the presence of toluene-p-sulfonic acid afforded two separable methyl hemiacetals 16a and 16b (Scheme 2). In order to derive 16a from 2, compound 2 was treated with MsCl and Et₃N as for the derivatives (12) of gymnastatin D 4. This reaction did not give the desired product, but compound 17 formed by rearrangement and aromatization (Scheme 2). The structure of 17 was established by analysis of ¹H-¹H COSY and HMBC correlations, and NOE data in 17. Cross peaks were observed from 2-H to 1-H, 3-H^{α} and 3-H^{β} in ¹H⁻¹H COSY, and from only one aromatic proton (5-H) to C-3 and two aromatic quaternary carbons (C-7 and C-9) bearing an oxygen atom, from 1-H to C-3 and C-9, and from $3-H^{\alpha}$ and $3-H^{\beta}$ to one (C-4) of three other aromatic quaternary carbons (C-4, C-6 and C-8) in HMBC. In additon to these correlations, the observation of NOEs from 5-H to $3-H^{\alpha}$ and $3-H^{\beta}$ suggested



Scheme 2 *Reaction conditions*: (a) Ac₂O, pyridine; (b) MsCl, Et₃N; (c) CH(OCH₃)₃, *p*-TsOH.

positional relationships of C-1–C-5, C-7 and C-9. Therefore, two chlorine atoms should be linked to the two remaining quaternary carbons (C-6 and C-8), and C-6 was assigned by an

	7a		
sition	$\delta_{\mathrm{H}}{}^{a}$	J/Hz	$\delta_{\rm C}$
	6.36d	4.4 (2)	95.3
	4.97dddd	$13.0(3\beta), 8.5(10), 8.0(3\alpha), 4.4(1)$	50.5
α	2.54dd	13.0 (3β), 8.0 (2)	37.9
β	1.96t	13.0 (2, 3α)	
			81.6
	5.74dd	2.5 (9), 1.8 (7)	129.6
			128.2
	5.69dd	2.8 (6), 1.8 (5)	66.5
	3.59dd	4.1 (9), 2.8 (7)	51.3
	3.43dd	4.1 (8), 2.5 (5)	55.5
	5.61d	8.5 (2)	
		(_)	166.2
	5.71d	14.7 (13)	116.3
	7.28d	14.7 (12)	148.0
	/1200	1, (12)	130.6
	5.69d	9.4 (16)	148.8
	2.51m		33.2
٨	1.26m		37.2

Table 6	¹ H and ¹³ C NMR data of gymnastatin D diacetates 7a and 7b in CDCl ₃
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	7a					7b			
Position	$\delta_{\mathbf{H}}{}^{a}$	J/Hz	$\delta_{ m C}$	HMBC (C)	NOESY	$\delta_{\rm H}{}^a$	<i>J</i> /Hz	$\delta_{\rm C}$	NOESY
1	6.36d	4.4 (2)	95.33 (t) ^b	2, 3, 4	2, 10	6.31s	4.4 (2)	100.85 (t)	2, 10
2	4.97dddd	13.0 (3β), 8.5 (10), 8.0 (3α), 4.4 (1)	50.56 (t)		1, 3α, 9, 10	4.69td	7.0 $(3\alpha, 10), 2.5 (3\beta)$	55.92 (t)	1, 3α, 9, 10
3α	2.54dd	13.0 (3β), 8.0 (2)	37.93 (s)	4, 5, 9	2, 3β, 9	2.63dd	14.1 (3β), 7.0 (2)	40.87 (s)	2,9
β	1.96t	13.0 (2, 3α)		1, 9	3α, 5	2.04dd	14.1 (3α), 2.5 (2)	~ /	5
4			81.65 (q)	2 -				83.52 (q)	
5	5.74dd	2.5 (9), 1.8 (7)	129.63 (t)	6, 7, 8	3β	5.82dd	2.5 (9), 1.8 (7)	129.11 (t)	3β
6			128.25 (q)	-, -, -			('),(')	128.28 (q)	- F
7	5.69dd	2.8 (6), 1.8 (5)	66.54 (t)	6	8	5.68dd	2.7 (8), 1.8 (5)	66.64 (t)	8
8	3.59dd	4.1 (9), 2.8 (7)	51.35 (t)	6, 7, 9	7,9	3.60dd	4.1 (9), 2.7 (7)	51.66 (t)	7,9
9	3.43dd	4.1 (8), 2.5 (5)	55.57 (t)	4	2, 3α, 8	3.43dd	4.1 (8), 2.5 (5)	56.67 (t)	2, 3α, 8
10	5.61d	8.5 (2)		2, 11	1, 2	5.78d	7.0 (2)		$1, 2, 3\beta$
11	biord	0.0 (2)	166.28 (q)	2, 11	1, 2	onou	/10 (2)	166.37 (q)	1, 2, 5 p
12	5.71d	14.7 (13)	116.33 (t)	11, 13, 14	23	5.75d	15.1 (13)	116.44(t)	23
13	7.28d	14.7 (12)	148.01 (t)	11, 12, 14, 15, 23	15	7.27d	15.1 (12)	147.86 (t)	15
14	7.20 u	11.7 (12)	130.66 (q)	11, 12, 11, 10, 20	15	7.27d	15.1 (12)	130.74 (q)	10
15	5.69d	9.4 (16)	148.88 (t)	13, 14, 16, 17, 23, 24	13, 24	5.68d	9.6 (16)	148.77 (t)	13, 24
16	2.51m	9.4 (10)	33.24(t)	15, 14, 10, 17, 25, 24	23, 24	2.50m	5.0 (10)	33.24 (t)	23, 24
17A	1.26m		37.20 (s)	18	23, 24	1.26m		37.22 (s)	23, 24
B	1.35m		57.20 (3)	18		1.34m		57.22 (3)	
18	1.22m		27.45 (s)	19		1.21m		27.46 (s)	
19	1.22m		29.36 (s)	19		1.23m		29.39 (s)	
20	1.23m		31.80 (s)	18		1.23m		31.81 (s)	
20	1.26m		22.60 (s)	19, 20		1.26m		22.62 (s)	
22	0.87t	6.9 (21)	14.07 (p)	20, 21		0.87t	6.9 (21)	14.09 (p)	
23	1.77s	0.9 (21)	12.50 (p)	13, 14, 15	12, 16	1.77s	0.9 (21)	12.51 (p)	12, 16
24	0.98d	6.7 (16)	20.50 (p)	15, 16, 17	12, 10	0.97d	6.6 (16)	20.52 (p)	15, 16
1-COCH ₃	0.700	0.7 (10)	169.13 (q)	10, 10, 17	10, 10	0. <i>7</i> / u	0.0 (10)	169.12 (g)	10, 10
1-COCH ₃	2.18s		21.37 (p)			2.18s		21.24 (p)	
7- <i>C</i> OCH ₃	2.105		170.37 (p)			2.105		170.35 (q)	
7-COCH ₃	2.20s		20.68 (p)			2.20s		20.70 (p)	
a,b As in Table			20.08 (p)			2.208		20.70 (p)	

Table 7 ¹ H and ¹³ C NMR data of gymnastatin B 2a in CD	Cl3
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Position	$\delta_{\mathrm{H}}{}^{a}$	J/Hz	$\delta_{ m C}$	¹ H– ¹ H COSY	HMBC	NOESY
1	4.65d	3.7 (2)	96.62 (t) ^{<i>b</i>}	2	1-OMe, 3	1-OMe, 2, 10
2	4.08dddd	$12.2(3\beta), 8.7(10), 4.7(3\alpha), 3.7(1)$	46.34 (t)	$1, 3\alpha, 10$,	1, 3α, 9
2 3α	2.23dd	12.2 (3β), 4.7 (2)	38.32 (s)	2, 3β	1, 2, 5, 9	2, 3β, 9
β	2.00t	12.2 (3α)		2, 3β	2, 3a	3α, 5
4			69.01 (q)	· •		, ,
5	4.29t	2.2 (6, 9)	75.46 (t)	6, 9	4, 6, 7, 9	1-OMe, 3β, 6
6	5.40d	2.2 (5)	60.85 (t)	5	5, 7	5, 5-OH
7			183.49 (q)		·	·
8			132.98 (q)			
9	6.94d	2.2 (5)	143.01 (t)	5	3, 5, 7, 8	3α
10	5.85d	8.7 (2)	~ ~ ~ ~ ~ ~ ~ ~ ~ ~ ~ ~ ~ ~ ~ ~ ~ ~ ~ ~	5 2	1, 2, 11	1, 12
11			166.59 (q)		, ,	,
12	5.72d	15.3 (13)	116.66 (t)	13	11, 13, 14	10, 23
13	7.14d	15.3 (12)	147.65 (t)	12	11, 12, 14, 15, 23	15
14			130.69 (q)		, , , , ,	
15	5.68d	9.6 (16)	148.91 (t)	16, 23	13, 14, 16, 17, 23, 24	13, 16, 24
16	2.51m		33.25 (t)	15, 17A, 17B, 24	, , , , , ,	15, 23, 24
17A	1.27m		37.14 (s)	16	18	, ,
В	1.34m			16	18	
18	1.22m		27.45 (s)		19	
19	1.24m		29.35 (s)			
20	1.27m		22.60 (s)		18	
21	1.23m		31.79 (s)	22	19, 20	
22	0.87t	6.8 (21)	14.05 (p)	21	20, 21	
23	1.76s		12.48 (p)	15	13, 14, 15	12, 16
24	0.97d	6.6 (16)	20.43 (p)	16	15, 16, 17	15, 16
1-OMe	3.47s	× ′	55.10 (p)		1	1, 5
5-OH	5.12br s		<i>u</i> ,			6
^{<i>a,b</i>} As in Tal	ble 1.					

HMBC correlation from 5-H to C-6. The *cis* arrangement of 1-H and 2-H was deduced from NOEs for 10-H/3-H^{β}, 10-H/ OMe, 1-H/2-H, 1-H/3-H^{α} and 2-H/3-H^{α}.

Since the above-mentioned reaction did not give the desired compound **16a**, compound **2** was next treated with acetic anhydride in pyridine. This reaction also afforded undesirable compounds, 5-acetate **18** and enol acetate **19**.

The absolute configuration for gymnastatin B 2 has not been established by a chemical transformation as described above, but is assumed to be the same as for its co-metabolites, gymnastatins A 1, D 4 and E 5.

Gymnastatin C **3** was assigned the molecular formula $C_{24}H_{35}Cl_2NO_5$ as deduced from an M⁺ peak of **3** in HREIMS. The general spectral features of compound **3** closely resembled those of **2** except that the trisubstituted double bond in the cyclohexane ring of **2** was replaced by two sp³-methines linked to a hydroxy group and a chlorine atom $[\delta_H 4.83 (8-H), \delta_C 66.58 (C-8); \delta_H 4.14 (9-H), \delta_C 73.80 (C-9)]$ in **3**, and the C-7 signal (δ_C 190.47) of the ketone in **3** appeared shifted downfield relative to that of **2** in the NMR spectra (Table 8). The planar structure for **3** established by analysis of the NMR spectra was confirmed by ¹H–¹H COSY and HMBC correlations (Table 8).

The observation of an NOE between 6-H and 8-H (Table 8) in 3 implied that the cyclohexane ring of 3 exists in a chair conformation with 6-H and 8-H in a coaxial arrangement. NOEs from 5-H to the 1-methoxy group, 6-H and 3-H^{β} , and the coupling constant ($J_{5,6}$ 3.6 Hz) indicated that the methoxy group and 3-H^{β} are on the same side as 5-H with an equatorial arrangement on the cyclohexane ring. The large coupling constant ($J_{8,9}$ 10.1 Hz) between 8-H and 9-H suggested that these protons are arranged *trans*-diaxially. In addition, NOEs from 3-H^{α} to 2-H and 9-H showed that 2-H is on the same side as 9-H and thus oriented *trans* to the 1-methoxy group. The summary of these considerations allowed assignment of relative stereostructure 3 to gymnastatin C.

Transformation from 3 to 16a for assignment of an absolute configuration to 3 was attempted by treatment with MsCl and

Et₃N. The resulting product was not the desired 16a, but the same compound 17 as obtained in the same reaction for 2. This result implied that the absolute configuration of C-1, C-2 and C-16 in 3 is the same as those of 2, and thus led to absolute stereostructure 3 for gymnastatin C.

Cytotoxic activities of compounds 1–5 were examined in the P388 lymphocytic leukemia test system in a cell culture, according to the method reported previously.²⁰ The results showed that three of the compounds (1–3) exhibited potent cytotoxic activity and two (4 and 5) exhibited weak cytotoxic activity (ED_{50} 0.018, 0.108, 0.106 and 10.5, 10.8 µg cm⁻³, respectively). Gymnastatin A 1 of these compounds showed strongest cytotoxicity. This evidence suggested that conjugated ketones were important for enhancement of cytotoxicity in gymnastatin analogues, and hence the cytotoxic activity of compound 3 resulted from a conjugated ketone which might be derived from compound 3 in the test system.

Experimental

General procedures

UV spectra were recorded on a Shimadzu spectrophotometer and IR spectra on a Perkin-Elmer FT-IR spectrometer 1720X. Optical rotations were obtained on a JASCO ORD/UV-5 spectropolarimeter and are given in units of $10^{-1} \text{ deg cm}^2 \text{ g}^{-1}$. CD spectra were recorded on a JASCO J-500A spectrometer. 1D and 2D NMR spectra were recorded at 27 °C on a Varian UNITY INOVA-500 spectrometer, operating at 500 and 125.7 MHz for ¹H and ¹³C, respectively, with TMS as an internal reference. EIMS was determined using a Hitachi M-4000H mass spectrometer. Liquid chromatography over silica gel (mesh 230-400) was performed under medium pressure. HPLC was run on a Waters ALC-200 instrument equipped with a differential refractometer (R 401) and Shim-pack PREP-ODS (25 cm × 20 mm i.d.). Analytical TLC was performed on precoated Merck aluminium sheets (DC-Alufolien Kieselgel 60 F254, 0.2 mm) with the solvent CH₂Cl₂-MeOH (19:1), and compounds were viewed under a UV lamp and sprayed with 10% H₂SO₄ followed by heating.

Table 8 ¹ H and ¹³ C NMR data of gymnastatin C 3 in CD	Cl ₃
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Position	$\delta_{ m H}{}^a$	J/Hz	δ_{C}	¹ H– ¹ H COSY	HMBC (C)	NOESY
1	4.67d	3.8 (2)	97.13 (t) ^b	2	1-OMe, 3	2, 10
2	4.22dddd	12.6 (3β), 8.7 (10), 5.2 (3α), 3.8 (1)	46.13 (t)	1, 3α, 3β, 10	1, 3, 11	1, 3α
3α	2.47dd	12.6 (3β), 5.2 (2)	34.66 (s)	3β	1, 2, 4, 9	2, 3β
β	1.73t	$12.6(2, 3\alpha)$		3α	1, 2, 4, 9	3a, 5
4			71.82 (q)			,
5	4.34d	3.6 (6)	74.02 (t)	6	3, 4, 6, 7, 9	1-OMe, 3β, 6
6	5.19d	3.6 (5)	61.21 (t)	5, 8	5, 7	5, 8
7		()	190.47 (g)	,	,	,
8	4.83d	10.1 (9)	66.58 (t)	6	3	6
9	4.14d	10.1 (8)	73.80 (t)	5, 8	7,9	3a, 5, 8
10	5.86d	8.7 (2)		,	1, 2, 11, 12	1, 12
11			166.61 (g)		7 7 7	,
12	5.71d	15.3 (13)	116.52 (t)	13	11, 13, 14	10, 23
13	7.20d	15.3 (12)	148.02 (t)	12	11, 12, 14, 15, 23	15
14			130.72 (q)		3 3 3 3 - 3 -	
15	5.68d	9.8 (16)	149.02 (t)	16, 23	13, 14, 16, 17, 23, 24	13, 16, 24
16	2.51m		33.26 (t)	15, 17A, 17B, 24		15, 23, 24
17A	1.26m		37.20 (s)	16	18	, ,
В	1.35m			16	18	
18	1.22m		27.47 (s)		19	
19	1.23m		29.38 (s)			
20	1.27m		22.62 (s)		18	
21	1.23m		31.82 (s)	22	19, 20	
22	0.87t	6.9 (21)	14.07 (p)	21	20, 21	
23	1.76s		12.49 (p)	15	13, 14, 15	12, 16
24	0.97d	6.6 (16)	20.47 (p)	16	15, 16, 17	15, 16
1-OMe	3.47s		55.27 (p)		1	5
5-OH	4.14br s		47			
9-OH	2.76br s					
^{<i>a,b</i>} As in Ta	ble 1.					

Culturing and isolation of metabolites

A strain of Gymnascella dankaliensis (Castellani) Currah OUPS-N134 was initially isolated from the sponge Halichondria japonica, collected in the Osaka Bay of Japan in April, 1994. The sponge was washed with EtOH and its slices applied to the surface of nutrient agar layered in a Petri dish. Serial transfers of one of the resulting colonies provided a pure strain of G. dankaliensis. The fungal strain was grown in a liquid medium (90 dm³) containing 1% malt extract, 1% glucose and 0.05% peptone in artificial seawater adjusted to pH 7.5 for four weeks at 27 °C. The culture was filtered under suction and the mycelia collected was extracted thrice with MeOH. The combined extracts were evaporated in vacuo to give a mixture of crude metabolites (11.0 g), the CH₂Cl₂-MeOH (1:1) soluble fraction of which exhibited cytotoxicity (ED₅₀ 21.5 μ g cm⁻³). The culture filtrate was extracted thrice with AcOEt. The combined extracts were evaporated in vacuo to afford a mixture of crude metabolites (1.6 g), the CH₂Cl₂-MeOH (1:1) soluble fraction of which exhibited cytotoxicity (ED_{50} 7.0 µg cm⁻³). The CH₂Cl₂-MeOH (1:1) soluble fraction of the MeOH extract was passed through Sephadex LH-20, using CH₂Cl₂-MeOH (1:1) as the eluent.

The second fraction (7.0 g), in which the activity was concentrated, was chromatographed on a silica gel column with a hexane-CH2Cl2-MeOH gradient as the eluent. The MeOH-CH₂Cl₂ (1:99) and (2:98) eluates were collected as 2 fractions [Fr. 1 (558.7 mg) and Fr. 2 (212.4 mg)] and 2 fractions [Fr. 3 (144 mg) and Fr. 4 (144.3 mg)], respectively. Silica gel column chromatography was repeated for Fr. 1 with a CH₂Cl₂-MeOH gradient as the eluent. The MeOH-CH₂Cl₂ (1:99) eluates were collected as 2 fractions [Fr. 5 (134.1 mg) and Fr. 6 (164.9 mg)], of which Fr. 6 was purified by HPLC using acetone-H₂O (4:1) to afford 3 fractions [Fr. 7 (6.7 mg), Fr. 8 (4.8 mg) and Fr. 9 (42.7 mg)]. Fr. 8 and Fr. 9 afforded 3 (1.5 mg), and 2 (11.1 mg) and 1 (19.6 mg), respectively, after purification by HPLC using acetone– H_2O (7:3) and (3:1) as the eluent, respectively. Fr. 3 and Fr. 4 were purified by HPLC using acetone– $H_2O(7:3)$ as the eluent to afford 4 (13.1 mg) and 5 (15.3 mg), respectively.

Gymnastatin A 1. Obtained as an amorphous powder, mp 74.2–76.0 °C, $[a]_D - 3.8$ ($c \ 0.73$, CHCl₃); λ_{max} (EtOH)/nm 266 (log ε /dm³ mol⁻¹ cm⁻¹ 4.63); ν_{max} (KBr)/cm⁻¹ 3377, 3277 (OH, NH), 1697 (C=C-CO), 1653 (CONH) and 1606 (C=C); m/z (EI) 437 (M⁺ - H₂O, 1.4%), 277 ([C₁₀H₁₁Cl₂NO₅]⁺, 53.8), 207 ([C₁₄H₂₃O]⁺, 72.7) and 178 ([C₁₃H₂₂]⁺, 100.0) [m/z (HREI) Found: M⁺ - H₂O, 437.1507. C₂₃H₂₉Cl₂NO₃ requires M - H₂O, 437.1523]; CD λ ($c \ 4.80 \times 10^{-5}$ mol dm⁻³ in EtOH)/nm 79 ($\Delta \varepsilon$ 0), 270 (+2.22), 267 (+2.02), 254 (+2.55), 244 (+2.49), 234 (+2.66) and 220 (0). This compound is a mixture of stereo-isomers **1a** and **1b** (2:1), and their ¹H and ¹³C NMR data are listed in Table 1.

Gymnastatin B 2. Obtained as an amorphous powder, mp 73.5–77.5 °C, $[a]_{\rm D}$ –122.1 (*c* 0.18, CHCl₃); $\lambda_{\rm max}$ (EtOH)/nm 265 (log ε /dm³ mol⁻¹ cm⁻¹ 4.50); $\nu_{\rm max}$ (KBr)/cm⁻¹ 3384, 3275 (OH, NH), 1725 (C=C-CO), 1650 (CONH) and 1610 (C=C); *m*/*z* (EI) 487 (M⁺, 2.0%), 277 ([C₁₀H₁₁Cl₂NO₅]⁺, 56.6), 207 ([C₁₄H₂₃O]⁺, 81.9), 178 ([C₁₃H₂₂]⁺, 100.0) and 95 ([C₅H₅NO]⁺, 37.2) [*m*/*z* (HREI) Found: M⁺, 487.1907. C₂₄H₃₅Cl₂NO₅ requires *M*, 487.1890]; CD λ (*c* 4.84 × 10⁻⁵ mol dm⁻³ in EtOH)/nm 286 ($\Delta\varepsilon$ 0), 258 (-5.63) and 234 (0). ¹H and ¹³C NMR data are listed in Table 7.

Gymnastatin C 3. Obtained as an amorphous powder, mp 104.7–107.5 °C, $[a]_{\rm D}$ –101.2 (*c* 0.12, CHCl₃); $\lambda_{\rm max}$ (EtOH)/nm 265 (log *e*/dm³ mol⁻¹ cm⁻¹ 4.42); $v_{\rm max}$ (KBr)/cm⁻¹ 3430 (OH, NH), 1755 (CO), 1651 (CONH) and 1604 (C=C); *m/z* (EI) 505 (M⁺, 1.0%), 487 ([M – H₂O]⁺, 2.3), 455 ([M – 2H₂O]⁺, 5.2), 277 ([C₁₀H₁₁Cl₂NO₅]⁺, 20.4), 207 ([C₁₄H₂₃O]⁺, 100.0), 179 ([C₁₃H₂₃]⁺, 68.7) and 95 ([C₅H₅NO]⁺, 80.6) [*m/z* (HREI) Found: M⁺, 505.1993. C₂₄H₃₇Cl₂NO₆ requires *M*, 505.1996]; CD λ (*c* 5.26 × 10⁻⁵ mol dm⁻³ in EtOH)/nm 356 ($\Delta \varepsilon$ 0), 328 (+0.72), 282 (0), 272 (+0.52), 263 (0) and 238 (-2.79). ¹H and ¹³C NMR data are listed in Table 8.

Gymnastatin D 4. Obtained as an amorphous powder, mp 86.4–88.2 °C, $[a]_D$ –8.9 (*c* 0.45, CHCl₃); λ_{max} (EtOH)/nm 265

(log ε /dm³ mol⁻¹ cm⁻¹ 4.44); v_{max} (KBr)/cm⁻¹ 3399, 3311 (OH, NH), 1652 (CONH) and 1612 (C=C); *m*/*z* (EI) 439 (M⁺, 1.4%), 421 ([M - H₂O]⁺, 14.0), 411 ([M - CO]⁺, 7.0), 326 ([M - C₈H₁₇]⁺, 18.0), 207 ([C₁₄H₂₃O]⁺, 100.0), 179 ([C₁₃H₂₃]⁺, 42.0) and 95 ([C₅H₅NO]⁺, 31.0) [*m*/*z* (HREI) Found: M⁺, 439.2127. C₂₃H₃₄CINO₅ requires *M*, 439.2124]; CD λ (*c* 5.16 × 10⁻⁵ mol dm⁻³ in EtOH)/nm 295 ($\Delta\varepsilon$ 0), 267 (-0.88), 241 (0), 220 (+1.70) and 215 (0). This compound is a mixture of stereoisomers **4a** and **4b** (2:1), and their ¹H and ¹³C NMR data are listed in Table 5.

Gymnastatin E 5. Obtained as an amorphous powder, mp 87.3–88.0 °C, $[a]_D - 8.5$ (*c* 0.52, CHCl₃); λ_{max} (EtOH)/nm 266 (log *e*/dm³ mol⁻¹ cm⁻¹ 4.63); ν_{max} (KBr)/cm⁻¹ 3334, 1652 (CONH) and 1612 (C=C); *m*/*z* (EI) 439 (M⁺, 1.4%), 326 ([M - C₈H₁₇]⁺, 16.0), 207 ([C₁₄H₂₃O]⁺, 100.0), 179 ([C₁₃H₂₃]⁺, 63.7) and 95 ([C₅H₅NO]⁺, 97.4) [*m*/*z* (HREI) Found: M⁺, 439.2122. C₂₃H₃₄ClNO₅ requires *M*, 439.2124]; CD λ (*c* 5.78 × 10⁻⁵ mol dm⁻³ in EtOH)/nm 288 (Δε 0), 272 (-0.39), 257 (0), 222 (+1.78) and 211 (0). This compound is a mixture of stereoisomers **5a** and **5b** (2:1), and their ¹H and ¹³C NMR data are listed in Table 3.

Acetylation of gymnastatin A 1

 Ac_2O (0.2 cm³) was added to a pyridine solution (0.2 cm³) of gymnastatin A 1 (23.3 mg), and the reaction mixture was left at room temperature overnight. The mixture was concentrated to dryness under reduced pressure, and the residue was purified by HPLC using acetone–H₂O (4:1) as the eluent to afford acetates **6a** (11.7 mg) and **6b** (5.3 mg).

Acetate 6a. Obtained as an amorphous powder, mp 175.5– 177.8 °C, $[a]_D - 19.7$ (*c* 0.91, CHCl₃); λ_{max} (EtOH)/nm 266 (log $\varepsilon/dm^3 mol^{-1} cm^{-1} 4.61$); ν_{max} (KBr)/cm⁻¹ 3313 (NH), 1756 (OCO), 1702 (C=C-CO), 1647 (CONH) and 1624 (C=C); *m/z* (EI) 501 (M⁺ + 4, 0.009%), 499 (M⁺ + 2, 0.053%), 497 (M⁺, 0.084%), 437 ([M - AcOH]⁺, 8.8), 207 ([C₁₄H₂₃O]⁺, 100.0), 179 ([C₁₃H₂₃]⁺, 59.7) and 95 ([C₅H₅NO]⁺, 61.2) [*m/z* (HREI) Found: M⁺, 497.1742. C₂₅H₃₃Cl₂NO₅ requires *M*, 497.1734]; CD λ (*c* 1.03 × 10⁻⁴ mol dm⁻³ in EtOH)/nm 337 ($\Delta \varepsilon$ 0), 292 (+0.74), 279 (0), 269 (-0.88), 262 (0), 245 (+2.75), 233 (0) and 218 (-3.22). ¹H and ¹³C NMR data are listed in Table 2.

Acetate 6b. Obtained as an amorphous powder, mp 68–69 °C, $[a]_{\rm D}$ –23.9 (*c* 0.53, CHCl₃); $v_{\rm max}$ (KBr)/cm⁻¹ 3269 (NH), 1751 (OCO), 1697 (C=C–CO), 1651 (CONH) and 1612 (C=C); *m/z* (EI) 497 (M⁺, 0.1%), 437 ([M – AcOH]⁺, 0.8), 207 ([C₁₄H₂₃O]⁺, 50.4), 179 ([C₁₃H₂₃]⁺, 56.1) and 95 ([C₅H₅NO]⁺, 100.0) [*m/z* (HREI) Found: M⁺, 497.1737. C₂₅H₃₃Cl₂NO₅ requires *M*, 497.1734]. ¹H and ¹³C NMR data are listed in Table 2.

Alkaline hydrolysis of acetate 6a

A solution (1 drop) of 28% ammonia in water was added to a solution of acetate **6a** (3.5 mg) in MeOH (2 cm³), and the reaction mixture was left at room temperature for 2 h. The solvent was evaporated off under reduced pressure, and the residue was purified by silica gel column chromatography with hexane–AcOEt (1:1) to afford compound **1** as a mixture of **1a** and **1b** (2:1).

Formation of keto-lactone 13 from gymnastatin A 1

A solution of gymnastatin A 1 (16 mg) in pyridine (0.2 cm³) was added to a pyridine–CrO₃ complex prepared from pyridine (0.2 cm³) and CrO₃ (50 mg), and the reaction mixture was left at room temperature overnight. The mixture was diluted with water and extracted with CH₂Cl₂. The extract was evaporated under reduced pressure, and the residue was subjected to silica gel column chromatography [CH₂Cl₂–MeOH (99:1)] followed

by HPLC [ODS; acetone-H₂O (4:1)] to afford keto-lactone 13 as an amorphous powder. Compound 13: mp 62.8-64.2 °C, [a]_D -127.9 (c 0.18, CHCl₃); v_{max} (KBr)/cm⁻¹ 3387, 3290 (NH), 1796 (γ -lactone), 1699 (C=C-CO), 1652 (CONH) and 1613 (C=C); m/z (EI) 455 ([M + H₂]⁺, 7.4%), 409 ([M - CO₂]⁺, 65.0), 294 ([M - CI₃H₂₃]⁺, 53.3), 207 ([CI₄H₂₃O]⁺, 89.4), 179 ([CI₃H₂₃]⁺, 84.8) and 95 ([C5H5NO]+, 100.0) [m/z (HREI) Found: M+, 455.1619. $C_{23}H_{31}Cl_2NO_4$ requires *M*, 455.1630]; δ_H (CDCl₃) $0.87 (3H, t, J = 6.9 Hz, 22-H_3), 0.98 (3H, d, J = 6.7 Hz, 24-H_3),$ 1.21 (2H, m, 18-H₂ or 19-H₂), 1.23 (4H, m, 19-H₂ or 18-H₂ and $21-H_2$ or $20-H_2$), 1.26 (3H, m, $20-H_2$ or $21-H_2$ and $17-H^{\beta}$), 1.35 (1H, m, 17-H^a), 1.77 (3H, s, 23-H₃), 2.51 (1H, m, 16-H), 2.61 (1H, dd, J = 13.5, 10.1 Hz, 3-H^{β}), 2.89 (1H, dd, J = 13.5, 10.1 Hz, $3-H^{\alpha}$), 4.63 (1H, td, J = 10.1, 6.0 Hz, 2-H), 5.71 (1H, d, *J* = 9.6 Hz, 15-H), 5.78 (1H, d, *J* = 15.3 Hz, 12-H), 6.16 (1H, br d, J = 6.0 Hz, 10-H), 7.07 (1H, d, J = 2.7 Hz, 9-H), 7.28 (1H, d, J = 2.7 Hz, 5-H) and 7.31 (1H, d, J = 15.3 Hz, 13-H); $\delta_{\rm C}$ (CDCl₃) 12.44 (C-23), 14.08 (C-22), 20.47 (C-24), 22.61 (C-20 or C-21), 27.45 (C-18 or C-19), 29.37 (C-19 or C-18), 31.80 (C-21 or C-20), 33.31 (C-16), 37.18 (C-17), 37.88 (C-3), 49.40 (C-2), 77.62 (C-4), 115.28 (C-12), 130.70 (C-14), 132.92 (C-8), 133.17 (C-6), 140.67 (C-9), 142.47 (C-5), 148.94 (C-13), 149.72 (C-15), 166.90 (C-11), 171.62 (C-7) and 172.27 (C-1); CD λ (c 5.23×10^{-5} mol dm⁻³ in EtOH)/nm 330 ($\Delta \epsilon$ 0), 262 (-19.35), 242 (0) and 223 (+7.04).

Methylation of gymnastatin A 1 by trimethyl orthoformate

A solution of gymnastatin A 1 (28 mg), toluene-*p*-sulfonic acid (3 mg) and trimethyl orthoformate (0.02 cm³) in anhydrous MeOH (3 cm³) was refluxed under argon for 2 h and then diluted with diethyl ether. It was washed with a 1: 1 mixture of 5% NaOH and saturated brine solutions, and then saturated brine solution, and evaporated under reduced pressure. The residue was purified by HPLC [acetone–H₂O (4:1)] to afford methyl acetals **16a** (6.9 mg) and **16b** (7.1 mg) as amorphous powders.

Methyl acetal 16a. Mp 119.5–120.5 °C, $[a]_D$ –31.4 (*c* 0.57, CHCl₃); v_{max} (KBr)/cm⁻¹ 3305 (NH), 1696 (C=C-CO), 1650 (CONH) and 1615 (C=C); m/z (EI) 469 (M⁺, 2.7%), 207 $([C_{14}H_{23}O]^+, 100.0), 179 ([C_{13}H_{23}]^+, 62.9) \text{ and } 95 ([C_5H_5NO]^+,$ 55.8) [m/z (HREI) Found: M⁺, 469.1778. C₂₄H₃₃Cl₂NO₄ requires M, 469.1786]; $\delta_{\rm H}$ (CDCl₃) 0.87 (3H, t, J = 6.9 Hz, 22-H₃), 0.98 (3H, d, J = 6.7 Hz, 24-H₃), 1.22 (2H, m, 18-H or 19-H), 1.23 (3H, m, 19-H or 18-H and 21-H or 20-H), 1.26 (1H, m, 17-H^β), 1.27 (2H, m, 20-H₂ or 21-H₂), 1.35 (1H, m, 17-H^α), 1.77 (3H, s, 23-H₃), 2.11 (1H, dd, J = 13.0, 11.2 Hz, 3-H^{β}), 2.51 $(1H, m, 16-H), 2.61 (1H, dd, J = 13.0, 8.2 Hz, 3-H^{\alpha}), 3.51 (3H, s, 3.5)$ 1-OMe), 4.83 (1H, dddd, J = 11.2, 8.6, 8.2, 4.5 Hz, 2-H), 5.01 (1H, d, J = 4.5 Hz, 1-H), 5.69 (1H, d, J = 9.8 Hz, 15-H), 5.75 (1H, d, J = 15.3 Hz, 12-H), 5.91 (1H, d, J = 8.6 Hz, 10-H), 6.95 (1H, d, J = 2.6 Hz, 5-H), 7.03 (1H, d, J = 2.6 Hz, 9-H) and 7.27 (1H, d, J = 15.3 Hz, 13-H); $\delta_{\rm C}$ (CDCl₃) 12.51 (C-23), 14.08 (C-22), 20.54 (C-24), 22.61 (C-20 or C-21), 27.46 (C-18 or C-19), 29.38 (C-19 or C-18), 31.82 (C-21 or C-20), 33.23 (C-16), 37.25 (C-17), 38.91 (C-3), 51.90 (C-2), 55.38 (1-OMe), 79.15 (C-4), 103.10 (C-1), 116.75 (C-12), 130.74 (C-14), 130.78 (C-8), 130.91 (C-6), 144.39 (C-9), 146.40 (C-5), 147.51 (C-13), 148.53 (C-15), 166.29 (C-11) and 172.39 (C-7).

Methyl acetal 16b. Mp 51.8–52.5 °C, $[a]_{\rm D}$ –28.8 (*c* 0.71, CHCl₃); $v_{\rm max}$ (KBr)/cm⁻¹ 3377, 3272 (NH), 1695 (C=C–CO), 1651 (CONH) and 1610 (C=C); *m*/*z* (EI) 469 (M⁺, 1.2%), 207 ([C₁₄H₂₃O]⁺, 100.0), 179 ([C₁₃H₂₃]⁺, 73.1) and 95 ([C₅H₅NO]⁺, 49.3) [*m*/*z* (HREI) Found: M⁺, 469.1774. C₂₄H₃₃Cl₂NO₄ requires *M*, 469.1786]; $\delta_{\rm H}$ (CDCl₃) 0.87 (3H, t, *J* = 6.8 Hz, 22-H₃), 0.98 (3H, d, *J* = 6.7 Hz, 24-H₃), 1.22 (2H, m, 18-H₂ or 19-H₂), 1.23 (2H, m, 21-H₂ or 20-H₂), 1.24 (2H, m, 19-H₂ or 18-H₂), 1.26 (3H, m, 17-H^β), 1.27 (2H, m, 20-H₂ or 21-H₂), 1.35

(1H, m, 17-H^a), 1.78 (3H, s, 23-H₃), 2.23 (1H, dd, J = 14.2, 1.6 Hz, 3-H^β), 2.51 (1H, m, 16-H), 2.72 (1H, dd, J = 14.2, 6.6 Hz, 3-H^a), 3.45 (3H, s, 1-OMe), 4.83 (1H, td, J = 6.6, 1.6 Hz, 2-H), 5.09 (1H, s, 1-H), 5.69 (1H, d, J = 9.8 Hz, 15-H), 5.75 (1H, d, J = 6.6 Hz, 10-H), 5.78 (1H, d, J = 15.3 Hz, 12-H), 6.99 (1H, d, J = 2.6 Hz, 5-H), 7.03 (1H, d, J = 2.6 Hz, 9-H) and 7.30 (1H, d, J = 15.3 Hz, 13-H); $\delta_{\rm C}$ (CDCl₃) 12.52 (C-23), 14.08 (C-22), 20.51 (C-24), 22.62 (C-20 or C-21), 27.46 (C-18 or C-19), 29.38 (C-19 or C-18), 31.81 (C-21 or C-20), 33.26 (C-16), 37.22 (C-17), 40.72 (C-3), 55.33 (1-OMe), 56.99 (C-2), 80.42 (C-4), 109.70 (C-1), 116.42 (C-12), 130.50 (C-8), 130.69 (C-14), 130.86 (C-6), 145.05 (C-5), 147.06 (C-9), 147.93 (C-13), 148.92 (C-15), 166.46 (C-11) and 172.45 (C-7).

Treatment of gymnastatin B 2 with MsCl and Et₃N

To a solution of gymnastatin B 2 (10.9 mg) in anhydrous CH₂Cl₂ were successively added Et₃N (0.05 cm³) and MsCl (0.025 cm³) under argon, while cooling the mixture in an ice bath. After stirring for 30 min, the reaction mixture was diluted with a 50% brine solution and extracted with CH₂Cl₂. The extract was washed with saturated aqueous NaHCO3 and water. After evaporation of the solvent, the residue was purified by HPLC [acetone-H₂O (4:1)] to afford 17 (6.9 mg) as an amorphous powder. Compound 17: mp 54.0-54.5 °C, [a]_D -25.9 (c 0.54, CHCl₃); v_{max} (KBr)/cm⁻¹ 3276 (NH), 1652 (CONH), 1615, 1538 (Ar C-C) and 1376, 1187 (OSO₂); m/z (EI) 547 (M⁺, 100%), 516 ([M - OCH₃]⁺, 11.3), 434 ([M - C₈H₁₇]⁺, 33.9), 381 ($[M - C_{12}H_{22}]^+$, 28.3), 324 ($[M - C_{14}H_{25}NO]^+$, 63.7), $293 ([M - C_{14}H_{25}NO - OCH_3]^+, 41.8), 207 ([C_{14}H_{23}O]^+, 64.3),$ 179 ($[C_{13}H_{23}]^+$, 53.1) and 95 ($[C_5H_5NO]^+$, 44.9) [*m*/*z* (HREI) Found: M⁺, 547.1564. C₂₅H₃₃Cl₂NO₆S requires *M*, 457.1560]; $\delta_{\rm H}$ (CDCl₃) 0.87 (3H, t, J = 6.6 Hz, 22-H₃), 0.98 (3H, d, J = 6.6Hz, 24-H₃), 1.22 (2H, m, 18-H₂ or 19-H₂), 1.23 (2H, m, 21-H₂ or 20-H₂), 1.24 (2H, m, 19-H₂ or 18-H₂), 1.26 (1H, m, 17-H^{β}), 1.27 (2H, m, 20-H₂ or 21-H₂), 1.35 (1H, m, 17-H^a), 1.78 (3H, s, 23-H₃), 2.51 (1H, m, 16-H), 2.82 (1H, ddd, J = 16.0, 12.0, 1.1 Hz, 3-H^{β}), 2.95 (1H, dd, J = 16.0, 6.1 Hz, 3-H^{α}), 3.47 (3H, s, 7-OSO₂Me), 3.52 (3H, s, 1-OMe), 4.57 (1H, dddd, J = 12.0, 8.9, 6.1, 2.5 Hz, 2-H), 5.19 (1H, d, J = 2.5 Hz, 1-H), 5.67 (1H, d, *J* = 8.9 Hz, 10-H), 5.68 (1H, d, *J* = 9.4 Hz, 15-H), 5.76 (1H, d, J = 15.3 Hz, 12-H), 7.09 (1H, s, 5-H) and 7.29 (1H, d, J = 15.3 Hz, 13-H); $\delta_{\rm C}$ (CDCl₃) 12.51 (C-23), 14.08 (C-22), 20.54 (C-24), 22.61 (C-20 or C-21), 27.00 (C-3), 27.46 (C-18 or C-19), 29.38 (C-19 or C-18), 31.82 (C-21 or C-20), 33.21 (C-16), 37.24 (C-17), 41.08 (7-OSO₂Me), 44.29 (C-2), 56.39 (1-OMe), 98.00 (C-1), 116.90 (C-12), 118.75 (C-6), 121.07 (C-8), 121.86 (C-4), 127.94 (C-5), 130.74 (C-14), 142.17 (C-7), 146.63 (C-9), 147.46 (C-13), 148.42 (C-15) and 166.17 (C-11); CD λ (c 4.94 × 10⁻⁵ mol dm⁻³ in EtOH)/nm 310 ($\Delta \varepsilon$ 0), 284 (-1.84), 268 (0), 245 (+0.92), 231 (0), 221 (+0.31), 217 (-0.25) and 206 (-3.31).

Treatment of gymnastatin C 3 with MsCl and Et₃N

Using the same procedure as above with compound **2**, gymnastatin C **3** (2.7 mg) was treated with Et_3N (0.05 cm³) and MsCl (0.025 cm³) in anhydrous CH₂Cl₂ and purified by HPLC [acetone–H₂O (4:1)] to afford compound **17** (0.8 mg) as an amorphous powder, identical with that derived from compound **2**.

Treatment of gymnastatin B 2 with Ac₂O

Using the same procedure as acetylation of comound 1, gymnastatin B 2 (5.7 mg) was treated with Ac₂O (0.1 cm³) in pyridine (0.1 cm³) and purified by HPLC [acetone–H₂O (4:1)] to afford 5-acetate 18 (2.3 mg) and enol acetate 19 (2.2 mg).

5-Acetate 18. Obtained as an amorphous powder, mp 73.0–73.5 °C, $[a]_D$ –83.5 (*c* 0.79, CHCl₃); v_{max} (KBr)/cm⁻¹ 3392, 3277 (NH), 1747 (OCO), 1731 (C=C–CO), 1650 (CONH) and 1611

(C=C); m/z (EI) 529 (M⁺, 14.3%), 469 ([M - Ac - OCH₃]⁺, 11.4), 277 ($[C_{10}H_{11}NO_5Cl_2]^+$, 28.1), 207 ($[C_{14}H_{23}O]^+$, 100.0), 179 ([C₁₃H₂₃]⁺, 79.3) and 95 ([C₅H₅NO]⁺, 71.3) [m/z (HREI) Found: M⁺, 529.2000. C₂₆H₃₇Cl₂NO₆ requires *M*, 529.1998]; $\delta_{\rm H}$ $(CDCl_3)$ 0.87 (3H, t, J = 6.9 Hz, 22-H₃), 0.97 (3H, d, J = 6.6 Hz, 24-H₃), 1.22 (2H, m, 18-H₂ or 19-H₂), 1.23 (2H, m, 21-H₂ or 20-H₂), 1.24 (2H, m, 19-H₂ or 18-H₂), 1.27 (3H, m, 20-H₂ or $21-H_2$ and $17-H^{\beta}$), 1.35 (1H, m, 17-H^{α}), 1.76 (3H, s, 23-H₃), 1.96 $(1H, t, J = 12.4 Hz, 3-H^{\beta})$, 2.07 (3H, s, 5-COMe), 2.50 (1H, m, 16-H), 3.03 (1H, dd, J = 12.4, 4.8 Hz, $3-H^{\alpha}$), 3.48 (3H, s, 1-OMe), 4.24 (1H, dddd, J = 12.4, 9.1, 4.8, 3.4 Hz, 2-H), 4.39 (1H, t, J = 2.3 Hz, 5-H), 4.66 (1H, d, J = 3.4 Hz, 1-H), 5.15 (1H, d, J = 2.3 Hz, 6-H), 5.57 (1H, d, J = 9.1 Hz, 10-H), 5.67 (1H, d, J = 9.8 Hz, 15-H), 5.69 (1H, d, J = 15.3 Hz, 12-H), 7.23 (1H, d, J = 15.3 Hz, 13-H) and 7.48 (1H, d, J = 2.3 Hz, 9-H); $\delta_{\rm C}$ (CDCl₃) 12.49 (C-23), 14.07 (C-22), 20.53 (C-24), 21.26 (5-COMe), 22.61 (C-20 or C-21), 27.45 (C-3), 27.45 (C-18 or C-19), 29.38 (C-19 or C-18), 31.81 (C-21 or C-20), 33.21 (C-16), 34.53 (C-3), 37.23 (C-17), 45.30 (C-2), 55.34 (1-OMe), 60.07 (C-6), 73.50 (C-5), 76.14 (C-4), 97.07 (C-1), 116.88 (C-12), 130.70 (C-14), 134.96 (C-8), 139.58 (C-9), 147.39 (C-13), 148.35 (C-15), 165.76 (C-11), 168.59 (5-COMe) and 182.11 (C-7).

Enol acetate 19. Obtained as an amorphous powder, mp 67.2–68.5 °C, [a]_D +85.4 (c 0.56, CHCl₃); v_{max} (KBr)/cm⁻¹ 3270 (NH), 1697 (C=C-CO), 1787, 1746 (OCO), 1651 (CONH) and 1613 (C=C); m/z (EI) 571 (M⁺, 0.9%), 511 ([M - OAc - H]⁺, 40.1), 246 ($[C_{10}H_9O_3Cl_2 - H]^+$, 100.0), 224 ($[C_{14}H_{24}NO + H]^+$, 59.9), 207 $([C_{14}H_{23}O]^+$, 42.1), 179 $([C_{13}H_{23}]^+$, 33.5) and 95 $([C_5H_5NO]^+, 31.7)$ [m/z (HREI) Found: M⁺, 571.2086. $C_{28}H_{39}Cl_2NO_7$ requires *M*, 571.2103]; δ_H (CDCl₃) 0.87 (3H, t, J = 6.9 Hz, 22-H₃), 0.97 (3H, d, J = 6.6 Hz, 24-H₃), 1.22 (2H, m, 18-H₂ or 19-H₂), 1.23 (2H, m, 19-H₂ or 18-H₂), 1.24 (2H, m, 21-H₂ or 20-H₂), 1.26 (1H, m, 17-H^β), 1.27 (2H, m, 20-H₂ or 21-H₂), 1.34 (1H, m, 17-H^a), 1.76 (3H, s, 23-H₃), 1.95 (1H, t, J = 12.4 Hz, $3-H^{\beta}$), 1.98 (3H, s, 5-COMe), 2.29 (3H, s, 7-COMe), 2.50 (1H, m, 16-H), 2.97 (1H, dd, J = 12.4, 3.6 Hz, 3- H^{α}), 3.51 (3H, s, 1-OMe), 4.21 (1H, d, J = 1.3 Hz, 5-H), 4.28 (1H, dddd, J = 12.4, 9.4, 3.6, 3.2 Hz, 2-H), 4.67 (1H, d, J = 3.2 Hz, 1-H), 5.51 (1H, d, J = 9.4 Hz, 10-H), 5.67 (1H, d, J = 9.9 Hz, 15-H), 5.69 (1H, d, J = 15.3 Hz, 12-H), 6.49 (1H, d, J = 1.3 Hz, 9-H) and 7.23 (1H, d, J = 15.3 Hz, 13-H); $\delta_{\rm C}$ (CDCl₃) 12.49 (C-23), 14.08 (C-22), 20.08 (7-COMe), 20.55 (C-24), 21.25 (5-COMe), 22.61 (C-20 or C-21), 27.45 (C-18 or C-19), 29.38 (C-19 or C-18), 31.81 (C-21 or C-20), 33.19 (C-16), 33.55 (C-3), 37.24 (C-17), 45.05 (C-2), 55.32 (1-OCH₃), 71.34 (C-5), 75.99 (C-4), 96.39 (C-1), 117.11 (C-12), 126.32 (C-9), 129.73 (C-6 and C-8), 130.73 (C-14), 140.75 (C-7), 147.15 (C-13), 148.10 (C-15), 165.75 (C-11), 166.76 (7-COMe) and 169.12 (5-COMe).

Acetylation of gymnastatin D 4

Using the same procedure as acetylation of compound 1, gymnastatin D 4 (19.7 mg) was treated with Ac₂O (0.2 cm³) in pyridine (0.2 cm³) and purified by HPLC using acetone–H₂O (7:3) as the eluent to afford diacetates 7a (12.3 mg) and 7b (2.9 mg) as an amorphous powder.

Diacetate 7a. Mp 74.5–75.0 °C, $[a]_D - 22.9$ (*c* 1.22, CHCl₃); v_{max} (KBr)/cm⁻¹ 3284 (NH), 1750 (OCO), 1655 (CONH) and 1615 (C=C); *m/z* (EI) 522 (M⁺, 1.7%), 463 ([M – OAc]⁺, 55.2), 404 ([M – 2OAc]⁺, 6.6), 350 ([M – 2OAc – C₄H₉]⁺, 29.6), 207 ([C₁₄H₂₃O]⁺, 100.0), 179 ([C₁₃H₂₃]⁺, 64.0) and 95 ([C₅H₅NO]⁺, 73.4) [*m/z* (HREI) Found: M⁺, 522.2241. C₂₇H₃₇ClNO₇ requires *M*, 522.2259]. ¹H and ¹³C NMR data are listed in Table 6.

Diacetate 7b. Mp 78.0–79.0 °C, $[a]_D$ –18.7 (*c* 0.30, CHCl₃); v_{max} (KBr)/cm⁻¹ 3274 (NH), 1749 (OCO), 1652 (CONH) and 1614 (C=C); *m*/*z* (EI) 522 (M⁺, 0.5%), 463 ([M – OAc]⁺, 100.0), 404 ([M – 2OAc]⁺, 12.8), 350 ([M – 2OAc – C₄H₃]⁺, 62.4),

206 ($[C_{14}H_{22}O]^+$, 100.0) and 177 ($[C_{13}H_{21}]^+$, 74.5) [*m*/*z* (HREI) Found: M⁺, 522.2247. C₂₇H₃₇ClNO₇ requires *M*, 522.2259]. ¹H and ¹³C NMR data are listed in Table 6.

Oxidation of gymnastatin D 4 by a pyridine–CrO₃ complex

Using the same procedure as above with compound 1, gymnastatin D 4 (29.1 mg) in pyridine (0.2 cm³) was treated with a pyridine-CrO₃ complex prepared from pyridine (0.2 cm³) and CrO₃ (50 mg) and purified by silica gel column chromatography [hexane-AcOEt (1:1)] followed by HPLC [ODS; acetone-H₂O (4:1)] to afford keto-lactone 11 (7.3 mg) as an amorphous powder. Keto-lactone 11: mp 80.5-81.5 °C, v_{max} (KBr)/cm⁻¹ 3365 (NH), 1795 (γ-lactone), 1713 (C=C-CO), 1652 (CONH) and 1615 (C=C); m/z (EI) 435 (M⁺) [m/z (HREI) Found: M⁺, 435.1818. C₂₃H₃₀ClNO₅ requires *M*, 435.1812]; $\delta_{\rm H}$ (acetone-d₆) 0.87 (3H, t, J = 6.9 Hz, 22-H₃), 0.99 (3H, d, J = 6.6 Hz, 24-H₃), 1.20-1.42 (10H, m, 17-21-H₂), 1.79 (3H, s, 23-H₃), 2.58 (1H, m, 16-H), 2.60 (1H, dd, J = 14.0, 8.9 Hz, 3-H^{β}), 3.10 (1H, dd, *J* = 14.0, 10.3 Hz, 3-H^a), 3.78 (1H, d, *J* = 4.1 Hz, 8-H), 4.24 (1H, dd, J = 4.1, 2.8 Hz, 9-H), 4.87 (1H, ddd, J = 10.3, 8.9, 7.1 Hz, 2-H), 5.70 (1H, d, J = 9.9 Hz, 15-H), 6.01 (1H, d, J = 15.3 Hz, 12-H), 7.14 (1H, d, J = 2.8 Hz, 6-H), 7.22 (1H, d, J = 15.3 Hz, 13-H) and 7.99 (1H, d, *J* = 7.1 Hz, 10-H).

Formation of chlorohydrin 12 from keto-lactone 11

A solution of keto-lactone 11 (3.2 mg) and pyridinium chloride (3.5 mg) in pyridine (0.2 cm^3) was left at room temperature overnight, and then diluted with water and extracted with CH₂Cl₂. The extract was washed with water and concentrated to dryness. The residue was purified by HPLC [acetone-H₂O (4:1)] to afford chlorohydrin 12 (5.9 mg) as an amorphous powder. Chlorohydrin 12: mp 81.5–82.5 °C, $[a]_D$ –17.9 (c 0.22, CHCl₃); v_{max} (KBr)/cm⁻¹ 3368, 3316 (NH, OH), 1790 (γlactone), 1723 (C=C-CO), 1651 (CONH) and 1615 (C=C); m/z (EI) 471 (M⁺, 2.0%), 453 ([M - H_2O]⁺, 32.5), 409 ([M - $CO_2]^+$, 22.4), 224 ([$C_{14}H_{26}NO$]⁺, 100.0), 207 ([$C_{14}H_{23}O$]⁺, 87.5), 179 ($[C_{13}H_{23}]^+$, 87.3) and 138 ($[C_{10}H_{18}]^+$, 83.7) [m/z (HREI) Found: M⁺, 471.1562. C₂₃H₃₁Cl₂NO₅ requires *M*, 471.1579]; $\delta_{\rm H}$ (CDCl₃) 0.87 (3H, t, J = 6.8 Hz, 22-H₃), 0.97 (3H, d, J = 6.6 Hz, 24-H₃), 1.20 (2H, m, 18-H₂ or 19-H₂), 1.23 (2H, m, 21-H₂ or 20-H₂), 1.24 (2H, m, 19-H₂ or 18-H₂), 1.26 $(1H, m, 17-H^{\beta})$, 1.27 (2H, m, 20-H₂ or 21-H₂), 1.34 (1H, m, 17-H^a), 1.76 (3H, s, 23-H₃), 2.51 (1H, m, 16-H), 2.53 (1H, dd, J = 13.1, 10.1 Hz, $3-H^{\beta}$), 3.05 (1H, dd, J = 13.1, 10.1 Hz, $3-H^{\alpha}$), 4.05 (1H, d, J = 11.0 Hz, 9-H), 4.73 (1H, td, J = 10.1, 6.1Hz, 2-H), 4.90 (1H, d, J = 11.0 Hz, 8-H), 5.69 (1H, d, J = 9.9 Hz, 15-H), 5.78 (1H, d, J=15.3 Hz, 12-H), 6.39 (1H, d, J = 6.1 Hz, 10-H), 7.13 (1H, s, 5-H) and 7.29 (1H, d, J = 15.3Hz, 13-H); $\delta_{\rm C}$ (CDCl₃) 12.50 (C-23), 14.09 (C-22), 20.45 (C-24), 22.62 (C-20 or C-21), 27.46 (C-18 or C-19), 29.37 (C-19 or C-18), 31.81 (C-21 or C20), 33.29 (C-16), 37.16 (C-17), 39.10 (C-3), 50.54 (C-2), 64.27 (C-8), 75.49 (C-9), 80.52 (C-4), 115.53 (C-12), 130.71 (C-14), 136.62 (C-6), 141.33 (C-5), 148.69 (C-13), 149.59 (C-15), 167.34 (C-11), 173.92 (C-1) and 182.64 (C-7).

Dehydration of chlorohydrin 12

(a). Using the same procedure as acetylation of gymnastatin A 1, chlorohydrin 12 (1.9 mg) was treated with Ac_2O (0.1 cm³) in pyridine (0.1 cm³) and purified by HPLC [acetone-H₂O (7:3)] to afford keto-lactone 13 (1.2 mg) as an amorphous powder, identical with that derived from gymnastatin A 1.

(b). Using the same procedure as above with gymnastatin B 2, chlorohydrin 12 (3.6 mg) was treated with Et_3N (0.05 cm³) and MsCl (0.05 cm³) in anhydrous CH₂Cl₂ (1 cm³) and purified by HPLC [acetone–H₂O (4:1)] to afford 13 (1.0 mg) as an amorphous powder.

Formation of the (*R*)- and (*S*)-MTPA esters 15a and 15b from gymnastatin D 4

(*R*)-MTPA (15 mg), dicyclohexylcarbodiimide (DCC) (20 mg) and 4-(dimethylamino)pyridine (DMAP) (10 mg) were added to a CH_2Cl_2 solution (1 cm³) of gymnastatin D **4** (7.9 mg), and the reaction mixture was left at room temperature for 3 h. The solvent was evaporated off under reduced pressure, and the residue was purified by silica gel column chromatography with hexane–AcOEt (1:1) followed by HPLC (ODS) using acetone– H_2O (4:1) as the eluent to afford ester **15a** (6.7 mg). The same reaction with **4** (5.5 mg) using (*S*)-MTPA (10 mg) gave ester **15b** (3.6 mg).

Ester 15a. Obtained as an amorphous powder; m/z (EI) 871 (M⁺) [m/z (HREI) Found: M⁺, 871.2923. C₄₃H₄₈ClF₆NO₉ requires M, 871.2922]; $\delta_{\rm H}$ (CDCl₃) 0.87 (3H, t, J = 6.9 Hz, 22-H₃), 0.99 (3H, d, J = 6.6 Hz, 24-H₃), 1.18–1.40 (10H, m, 17– 21-H₂), 1.75 (3H, s, 23-H₃), 1.85 (1H, t, J = 13.0 Hz, 3-H^β), 2.43 (1H, dd, J = 13.0, 8.0 Hz, 3-H^α), 2.52 (1H, m, 16-H), 3.43 (1H, dd, J = 4.1, 2.7 Hz, 9-H), 3.53 (3H, s, OMe), 3.62 (3H, s, OMe), 3.72 (1H, dd, J = 4.1, 2.8 Hz, 6-H), 5.07 (1H, dddd, J = 13.0, 8.9, 8.0, 4.3 Hz, 2-H), 5.38 (1H, dd, J = 2.8, 1.4 Hz, 5-H), 5.40 (1H, d, J = 8.9 Hz, 10-H), 5.73 (1H, dd, J = 2.7, 1.4 Hz, 7-H), 6.54 (1H, d, J = 4.3 Hz, 1-H), 7.26 (1H, d, J = 15.3 Hz, 13-H), 7.42 (6H, m, ArH), 7.49 (2H, m, ArH) and 7.60 (2H, m, ArH).

Ester 15b. Obtained as an amorphous powder; m/z (EI) 871 (M⁺) [m/z (HREI) Found: M⁺, 871.2926. C₄₃H₄₈ClF₆NO₉ requires M, 871.2922]; $\delta_{\rm H}$ (CDCl₃) 0.87 (3H, t, J = 6.9 Hz, 22-H₃), 0.99 (3H, d, J = 6.6 Hz, 24-H₃), 1.18–1.40 (10H, m, 17–21-H₂), 1.77 (3H, s, 23-H₃), 1.80 (1H, t, J = 13.1 Hz, 3-H^β), 2.46 (1H, dd, J = 13.1, 8.0 Hz, 3-H^α), 2.52 (1H, m, 16-H), 3.39 (1H, dd, J = 4.1, 2.5 Hz, 9-H), 3.53 (3H, s, OMe), 3.61 (3H, s, OMe), 3.64 (1H, dd, J = 4.1, 3.0 Hz, 8-H), 5.06 (1H, dddd, J = 13.1, 8.9, 8.0, 4.1 Hz, 2-H), 5.20 (1H, d, J = 8.9 Hz, 10-H), 5.42 (1H, d, J = 15.3 Hz, 12-H), 5.54 (1H, dd, J = 3.0, 1.6 Hz, 7-H), 6.59 (1H, d, J = 4.1 Hz, 1-H), 7.26 (1H, d, J = 15.3 Hz, 13-H), 7.42 (3H, m, ArH), 7.47 (3H, m, ArH), 7.52 (2H, m, ArH) and 7.58 (2H, m, ArH).

Acetylation of gymnastatin E 5

Using the same procedure as above with compound 1, gymnastatin E 5 (15.8 mg) was treated with Ac₂O (0.2 cm³) in pyridine (0.2 cm³) and purified by HPLC using acetone–H₂O (7:3) as the eluent to afford diacetates 8a (12.7 mg) and 8b (2.7 mg).

Diacetate 8a. Obtained as colourless plates, mp 100.5–103.0 °C, $[a]_D$ -36.4 (*c* 0.31, CHCl₃); v_{max} (KBr)/cm⁻¹ 3283 (NH), 1748 (OCO), 1657 (CONH) and 1614 (C=C); *m/z* (EI) 522 (M⁺, 0.5%), 463 ([M – OAc]⁺, 27.5), 404 ([M – 2OAc]⁺, 11.1), 350 ([M – 2OAc – C₄H₃]⁺, 13.6), 207 ([C₁₄H₂₃O]⁺, 100.0) and 179 ([C₁₃H₂₃]⁺, 74.5) [*m/z* (HREI) Found: M⁺, 522.2244. C₂₇H₃₇ClNO₇ requires *M*, 522.2259]. ¹H and ¹³C NMR data are listed in Table 4.

Diacetate 8b. Obtained as an amorphous powder, mp 67.0– 68.0 °C, $[a]_D$ –15.4 (*c* 0.27, CHCl₃); v_{max} (KBr)/cm⁻¹ 3267 (NH), 1751 (OCO), 1651 (CONH) and 1613 (C=C); *m/z* (EI) 522 (M⁺, 1.1%), 463 ([M – OAc]⁺, 100.0), 404 ([M – 2OAc]⁺, 80.7), 350 ([M – 2OAc – C₄H₉]⁺, 66.1), 207 ([C₁₄H₂₃O]⁺, 97.6) and 177 ([C₁₃H₂₁]⁺, 74.5) [*m/z* (HREI) Found: M⁺, 522.2223. C₂₇H₃₇ClNO₇ requires *M*, 522.2259]. ¹H and ¹³C NMR data are listed in Table 4.

Oxidation of gymnastatin E 5 by a pyridine–CrO₃ complex

Using the same procedure as above with compound 1,

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gymnastatin E 5 (25.5 mg) in pyridine (0.2 cm³) was treated with a pyridine– CrO_3 complex prepared from pyridine (0.2 cm³) and CrO₃ (50 mg) and purified by silica gel column chromatography [hexane-AcOEt (1:1)] followed by HPLC [acetone- $H_2O(4:1)$] to afford keto-lactone 9 (3.8 mg) as an amorphous powder. Keto-lactone 9: mp 81.0-82.0 °C, [a]_D -52.4 (c 0.37, CHCl₃); ν_{max} (KBr)/cm⁻¹ 3294 (NH), 1795 (γ-lactone), 1713 (C=C-CO), 1653 (CONH) and 1614 (C=C); m/z (EI) 435 (M⁺, 5.1%), 322 ($[M - C_8H_{17}]^+$, 8.2), 224 ($[C_{14}H_{26}NO]^+$, 100.0), 207 $([C_{14}H_{23}O]^+, 31.6), 179 ([C_{13}H_{23}]^+, 50.6) \text{ and } 95 ([C_5H_5NO]^+,$ 76.2) [m/z (HREI) Found: M⁺, 435.1812. C₂₃H₃₀ClNO₅ requires *M*, 435.1812]; $\delta_{\rm H}$ (CDCl₃) 0.87 (3H, t, *J* = 6.9 Hz, 22-H₃), 0.97 (3H, d, J = 6.6 Hz, 24-H₃), 1.20 (2H, m, 18-H₂ or 19-H₂), 1.23 (4H, m, 19-H₂ or 18-H₂ and 21-H₂ or 20-H₂), 1.26 (1H, m, $17-H^{\beta}$), 1.27 (2H, m, 20-H₂ or 21-H₂), 1.35 (1H, m, 18-H^{α}), 1.76 (3H, s, 23-H₃), 2.51 (1H, m, 16-H), 2.64 (1H, dd, J = 13.5, 10.0 Hz, $3-H^{\beta}$), 2.77 (1H, dd, J = 13.5, 10.0 Hz, $3-H^{\alpha}$), 3.70 (1H, d, *J* = 3.9 Hz, 6-H), 4.12 (1H, dd, *J* = 3.9, 2.8 Hz, 5-H), 4.62 (1H, td, J = 10.0, 6.3 Hz, 2-H), 5.69 (1H, d, J = 9.8 Hz, 15-H), 5.78 (1H, d, J = 15.2 Hz, 12-H), 6.31 (1H, d, J = 6.3 Hz, 10-H), 6.72 (1H, d, *J* = 2.8 Hz, 9-H) and 7.29 (1H, d, *J* = 15.2 Hz, 13-H); $\delta_{\rm C}$ (CDCl₃) 12.44 (C-23), 14.08 (C-22), 20.46 (C-24), 22.61 (C-20 or C-21), 27.45 (C-18 or C-19), 29.37 (C-19 or C-18), 31.80 (C-21 or C-20), 33.30 (C-16), 37.17 (C-17), 37.92 (C-3), 49.38 (C-2), 51.90 (C-6), 55.66 (C-5), 79.79 (C-4), 115.38 (C-12), 129.66 (C-8), 130.72 (C-14), 139.48 (C-9), 148.82 (C-13), 149.63 (C-15), 166.96 (C-11), 172.70 (C-1) and 185.52 (C-7).

Formation of chlorohydrin 10 from keto-lactone 9

Using the same procedure as above with compound 11, ketolactone 9 (3.2 mg) was treated with pyridinium chloride (3.5 mg) in pyridine (0.2 cm³) and purified by HPLC [acetone-H₂O (3:1)] to afford chlorohydrin 10 (1.9 mg) as an amorphous powder. Chlorohydrin 10: mp 70.0-71.0 °C, [a]_D -42.1 (c 0.19, CHCl₃); ν_{max} (KBr)/cm⁻¹ 3359, 3302 (NH, OH), 1790 (γlactone), 1723 (C=C-CO), 1652 (CONH) and 1614 (C=C); m/z (EI) 471 (M⁺, 0.8%), 453 ([M - H_2O]⁺, 14.7), 409 ([M -CO₂]⁺, 35.1), 224 ([C₁₄H₂₆NO]⁺, 100.0), 207 ([C₁₄H₂₃O]⁺, 94.4), 179 ($[C_{13}H_{23}]^+$, 90.1), 138 ($[C_{10}H_{18}]^+$, 53.8) and 95 ($[C_5H_5NO]^+$, 32.5) [*m*/*z* (HREI) Found: M⁺, 471.1571. C₂₃H₃₁Cl₂NO₅ requires M, 471.1579]; $\delta_{\rm H}$ (CDCl₃) 0.87 (3H, t, J=6.9 Hz, 22-H₃), 0.98 (3H, d, J = 6.6 Hz, 24-H₃), 1.22 (2H, m, 18-H₂ or 19-H₂), 1.23 (2H, m, 21-H₂ or 20-H₂), 1.24 (2H, m, 19-H₂ or 18-H₂), 1.27 (3H, m, 17-H^{β} and 20-H₂ or 21-H₂), 1.36 (1H, m, 18-H^α), 1.76 (3H, s, 23-H₃), 2.51 (1H, m, 16-H), 2.68 (1H, dd, J = 13.9, 8.2 Hz, $3 \cdot H^{\beta}$), 2.84 (1H, dd, J = 13.9, 10.1 Hz, $3 \cdot H^{\alpha}$), 3.62 (1H, br s, 5-OH), 4.05 (1H, d, J = 10.4 Hz, 5-H), 4.92 (1H, d, J = 10.4 Hz, 6-H), 5.05 (1H, ddd, J = 10.1, 8.9, 8.2 Hz, 2-H), 5.69 (1H, d, J = 9.9 Hz, 15-H), 5.77 (1H, d, J = 15.3 Hz, 12-H), 6.34 (1H, d, J = 8.9 Hz, 10-H), 7.03 (1H, s, 9-H) and 7.30 (1H, d, J = 15.3 Hz, 13-H); δ_{C} (CDCl₃) 12.45 (C-23), 14.09 (C-22), 20.49 (C-24), 22.62 (C-20 or C-21), 27.46 (C-18 or C-19), 29.38 (C-19 or C-18), 31.82 (C-21 or C-20), 33.28 (C-16), 37.20 (C-17), 37.46 (C-3), 48.27 (C-2), 63.76 (C-6), 74.53 (C-5), 80.61 (C-4), 115.85 (C-12), 130.76 (C-14), 136.35 (C-8), 139.50 (C-9), 148.56 (C-13), 149.29 (C-15), 166.81 (C-11), 172.88 (C-1) and 182.13 (C-7).

Dehydration of chlorohydrin 10 by Ac₂O

Using the same procedure as acetylation of gymnastatin A 1, chlorohydrin 10 (1.1 mg) was treated with Ac₂O (0.1 cm³) in pyridine (0.1 cm³) and purified by HPLC [acetone–H₂O (4:1)] to afford keto-lactone 13 (0.8 mg) as an amorphous powder, identical with that derived from gymnastatin A 1.

Formation of the (R)- and (S)-MTPA esters 14a and 14b from gymnastatin E 5

Using the same procedure as above with gymnastatin D 4,

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gymnastatin E 5 (5.3 and 3.7 mg) was treated with (R)-MTPA (10 mg) and (S)-MTPA (10 mg) to afford esters 14a (4.1 mg) and 14b (4.3 mg), respectively.

Ester 14a. Obtained as an amorphous powder; m/z (EI) 871 (M⁺) [m/z (HREI) Found: M⁺, 871.2919. C₄₃H₄₈ClF₆NO₉ requires M, 871.2922]; $\delta_{\rm H}$ (CDCl₃) 0.87 (3H, t, J = 6.9 Hz, 22-H₃), 0.99 (3H, d, J = 6.7 Hz, 24-H₃), 1.18–1.38 (10H, m, 17–21-H₂), 1.74 (3H, s, 23-H₃), 1.94 (1H, t, J = 12.6 Hz, 3-H^β), 2.34 (1H, dd, J = 12.6, 8.2 Hz, 3-H^α), 2.52 (1H, m, 16-H), 2.94 (1H, dd, J = 3.9, 2.4 Hz, 5-H), 3.51 (1H, dd, J = 3.9, 2.8 Hz, 6-H), 3.51 (3H, s, OMe), 3.60 (3H, s, OMe), 5.03 (1H, dddd, J = 12.6, 9.0, 8.2, 4.3 Hz, 2-H), 5.42 (1H, d, J = 9.0 Hz, 10-H), 5.55 (1H, d, J = 15.2 Hz, 12-H), 5.68 (1H, d, J = 9.3 Hz, 15-H), 5.76 (1H, dd, J = 2.8, 1.6 Hz, 7-H), 5.90 (1H, dd, J = 2.4, 1.6 Hz, 9-H), 6.53 (1H, d, J = 4.3 Hz, 1-H), 7.26 (1H, d, J = 15.2Hz, 13-H), 7.39 (3H, m, ArH), 7.43 (3H, m, ArH), 7.48 (2H, m, ArH) and 7.59 (2H, m, ArH).

Ester 14b. Obtained as an amorphous powder; m/z (EI) 871 (M⁺) [m/z (HREI) Found: M⁺, 871.2920. C₄₃H₄₈ClF₆NO₉ requires M, 871.2922]; $\delta_{\rm H}$ (CDCl₃) 0.87 (3H, t, J = 6.9 Hz, 22-H₃), 0.99 (3H, d, J = 6.6 Hz, 24-H₃), 1.19–1.41 (10H, m, 17–21-H₂), 1.77 (3H, s, 23-H₃), 1.89 (1H, t, J = 12.8 Hz, 3-H^β), 2.39 (1H, dd, J = 12.8, 8.5 Hz, 3-H^α), 2.52 (1H, m, 16-H), 3.18 (1H, dd, J = 4.0, 2.6 Hz, 5-H), 3.55 (3H, s, OMe), 3.63 (3H, s, OMe), 3.67 (1H, dd, J = 4.0, 3.0 Hz, 6-H), 5.05 (1H, dddd, J = 12.8, 8.9, 8.5, 4.4 Hz, 2-H), 5.19 (1H, d, J = 8.9 Hz, 10-H), 5.42 (1H, d, J = 15.1 Hz, 12-H), 5.68 (1H, d, J = 2.6, 1.8 Hz, 9-H), 6.63 (1H, d, J = 4.4 Hz, 1-H), 7.22 (1H, d, J = 15.1 Hz, 13-H), 7.42 (3H, m, ArH), 7.45 (3H, m, ArH), 7.54 (2H, m, ArH) and 7.60 (2H, m, ArH).

X-Ray crystallography of diacetate 8a from gymnastatin E 5

Diacetate 8a from gymnastatin E 5 was crystallized from methanol solution by the vapour diffusion method. Crystal data: $C_{27}H_{37}CINO_{7}$, M = 523.03, monoclinic, C2, a = 19.175(7) Å, b = 9.322(3) Å, c = 17.751(4) Å, $\beta = 103.00(2)^{\circ}$, V = 3091.8(15) Å³, Z = 4, $d_x = 1.124$ Mg m⁻³, F(000) = 1116, μ (Cu-Ka) = 1.422 cm⁻¹. Data collection was performed by a Rigaku AFC5R using graphite-monochromated radiation ($\lambda = 1.5418$ Å). Total 2743 reflections were collected until $\theta = 63.29^{\circ}$, in which 2557 reflections were observed $[I > 2\sigma(I)]$. The crystal structure was solved by the direct method using SHELXS-86.²¹ The structure was refined by the full matrix least-squares method on F using SHELXL-93.22 In the structure refinements, non-hydrogen atoms were refined with anisotropic temperature factors. Hydrogen atoms were calculated on the geometrically ideal positions by the 'ride on' method, and were included in the calculation of structure factors with isotropic temperature factors. In the final stage, R = 0.0882, Rw = 0.2259 and S = 1.061were obtained.

Full crystallographic details, excluding structure factor tables, have been deposited at the Cambridge Crystallographic Data Centre (CCDC). For details of the deposition scheme, see 'Instructions for Authors', *J. Chem. Soc.*, *Perkin Trans. 1*, available *via* the RSC Web page (http://www.rsc.org/authors). Any request to the CCDC for this material should quote the full literature citation and the reference number 207/262.

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